



Research Article

# Evaluation of a Few Biomarkers among COVID-19 Patients

Rusul Heider Mohsin<sup>1</sup>, Deemah Saeb Abdul Hussein<sup>1</sup>, Ahmed S Abed<sup>2</sup>

<sup>1</sup>Department of Anesthesia, Hilla University Collage, Babylon, Iraq.

<sup>2</sup>Department of Medical Physics, Hilla University Collage, Babylon, Iraq.

DOI: <https://doi.org/10.24321/0019.5138.202213>

## I N F O

### Corresponding Author:

Ahmed S Abed, Department of Medical Physics,  
Hilla University Collage, Babylon, Iraq.

### E-mail Id:

a77medsalim2@gmail.com

### Orcid Id:

<https://orcid.org/0000-0001-6826-1942>

### How to cite this article:

Mohsin RH, Hussein DSA, Abed AS. Evaluation of a Few Biomarkers among COVID-19 Patients. Special Issue - COVID-19 & Other Communicable Disease. 2022;82-90.

Date of Submission: 2021-10-26

Date of Acceptance: 2021-12-22

## A B S T R A C T

**Introduction:** In the present study, the mean differences of Lactate Dehydrogenase (LDH), D-dimer, C-reactive protein, interleukin-6 and ferritin levels concentrations between study groups (patients with severe COVID-19 symptoms and patients with mild symptoms) compared to the control group. The results showed that there were significant increased concentrations of biomarkers levels in group A as compared with group B and control.

**Objective:** This present study aims to evaluate the COVID-19 biomarkers (Lactate Dehydrogenase (LDH), D-dimer, C-reactive protein, IL-6 and ferritin concentrations) among COVID-19 patients.

**Methodology:** A total of 75 blood samples were collected from male patients with age groups ranging between 30 and 75 years who were suffering from coronavirus. Three groups were included in this study; each group includes 25 patients. Group A patients suffering from coronavirus with severe symptoms, group B patients suffering from coronavirus with mild symptoms and group C with healthy patients as control. All parameters were measured according to standard procedures. Data were analysed in SPSS version 20 by using mean  $\pm$  SD. Significant association was established by chi-square test taking  $p$ -value  $< 0.05$ .

**Results:** Increased LDH values were linked to an increase in COVID-19 toxicity. On the basis of D-dimer, the probability of mortality can be determined. C-responsive protein and ferritin serum exercises were significantly increased in COVID-19 patients compared to those with mild side effects of COVID-19. IL-6 is a key immunomodulatory cytokine in both normal and infected tissues.

**Conclusion:** LDH, D-dimer, C-reactive protein and serum ferritin are good predictors of COVID-19 severity and may be used for the assessment of clinical outcome.

**Keywords:** COVID-19, Lactate Dehydrogenase (LDH), IL-6, D-dimer, C-reactive Protein, Ferritin



## Introduction

The coronavirus disease 2019 (COVID-19) is a profoundly irresistible sickness where the primary tainted case was revealed in Wuhan city-China from where it spread around the world.<sup>1</sup> The causative operator has a place with novel encompassed single straight positive-sense abandoned RNA Coronavirus, which is likewise called SARS-CoV-2 and has a fondness for lung cells.<sup>2</sup> The hereditary examination of SARS-CoV-2 proposed that this novelty HGN I strain might be created from the creature. Root by recombination between a bat SARS-like CoV and a coronavirus of obscure birthplace.<sup>3</sup> The capacity of fast spread of SARS-CoV-2 infection from individual to other is comparable or much more than to other human infections like flu or plague prompting to be declared as a pandemic by WHO in 2020.<sup>4</sup> The realities in this audit lead to propose that as a rule, the demise in SARS-CoV-2 may happen through loss of fundamental fiery reaction control which prompts lung injury followed by pneumonia, intense respiratory misery condition (ARDS) and respiratory disappointment, thus passing particularly in old patients with incessant infection.<sup>5</sup> Individuals with hypertension are likewise marginally bound to bite the dust from coronavirus. They are at a 6% higher risk than the general populace.<sup>6</sup> Numerous patients with extreme COVID-19 are those with coinciding, constant conditions, including hypertension and diabetes.<sup>7</sup> Both of these conditions expands the danger of kidney infection. Like other respiratory malady prompting infections, for example, SARS, MERS, and network gained pneumonia (CAP), COVID-19 typically first influences the lungs. Early side effects incorporate fever, cold, restlessness and difficulty in breathing.<sup>8</sup> These side effects can show up within two days or as long as 14 days after contact with the infection.<sup>9</sup> The COVID-19 seriousness shifts from mellow or asymptomatic to extreme or deadly. More established individuals and those with interminable maladies have all the earmarks of being at a higher danger of genuine manifestations.<sup>10</sup> This fluctuation is additionally found in the impact of COVID-19 on the lungs.<sup>11</sup> A few people may have just mellow respiratory side effects while others may create pneumonia, which doesn't compromise an individual's life.<sup>12</sup> Few people additionally experience the ill effects of extreme lung problems. As indicated by look into, what we regularly find in individuals who are seriously contaminated with COVID-19 is respiratory pain disorder.<sup>13</sup> Non-specific symptoms admire fever, fatigue, cough (with or while not liquid body substance production), anorexia, malaise, muscle pain, sore throat, dyspnea, nasal congestion, or headache might occur in patients with uncomplicated higher tract infection; seldom can there be diarrhoea, fatigue, and disgorgement in patients too. Grownup and immunological disorder patients with uncommon symptoms will show up.<sup>14</sup> Symptoms from pregnancy physiological changes or adverse effects during pregnancy (e.g., dyspnoea, fever,

gastrointestinal indications, weariness) can relate to this disease.<sup>15</sup> Pneumonia without signs of severe pneumonia and no requirement for more oxygen.<sup>16</sup>

## Materials and Method

A total of 75 blood samples were collected from male patients with age group range between (30-75 years) who were suffering from coronavirus. Samples were collected from patients during the period from January 2021 to March 2021 from the Marjan Medical City and private laboratories in Hilla, Iraq.

## Study Groups

Three groups were included in this study; each group includes 25 patients. Group A patients suffering from coronavirus with severe symptoms, group B patients suffering from coronavirus with mild symptoms and group C with healthy patients as control. All patients and control were from the same ethnic group (Arabic).

## Collection of Blood Samples

5 ml of blood was collected from each subject by vein puncture method. 2 mL of blood was kept into EDTA vials with sodium citrate for separating plasma for the estimation of D-dimer whereas, 3 mL of blood was kept into dispensable tubes containing isolating gel, and was allowed to clot at room temperature for 30 minutes. After that, it was centrifuged at 2000 rpm for more or less 10 min for serum separation. The serum was transferred into little aliquots and stored at (-20°C) for the estimation of other biomarkers.

## Determination of LDH Level Concentration

Estimation of LDH level in serum was done by using an auto analyzer for spectrometry and measured by mind ray system, China.

## Determination of D-dimer Level Concentration

Estimation of D-dimer level was done using a fluorescence immunoassay that quantifies the total D-dimer concentration in plasma. The test was used as an aid in the post-therapeutic evaluation of thromboembolic disease patients. A specific kit from Boditech Bio-technology, France was also used for the estimation.

## Determination of C-reactive Protein Concentration

In most cases, the blood is taken from a vein in the arm. Using a rubber band to wrap around the arm, blood is drawn from the veins and the puncture site is disinfected. A vial or syringe can be used to collect a small amount of blood. Afterwards, the bandage is removed to allow blood to flow freely again. The needle is removed and the puncture site is sealed with a crushed membrane once enough blood has been extracted. Only a few minutes are needed for this rather easy procedure. Mispa2, assessed C-reactive protein using an auto-analyzers system, Switzerland.

**Determination of IL-6 Concentration**

The human IL-6enzyme-linked immunosorbent assay kit was used in this study for the quantitative determination of cytokine concentration in adult patients’ serum samples and was done according to company instruction (Elabscience, China).

**Determination of Ferritin Level Concentration**

Estimation of ferritin level concentration in serum was done by an auto-analyzer for spectrometry and measured by cobase e 411, Germany.

**Statistical Analysis**

After collection of data, it was entered in Microsoft excel sheet. Data were analysed in SPSS version 20 by using mean ±SD. Significant association was established by chi-square test taking p-value<0.05.<sup>17</sup>

**Results**

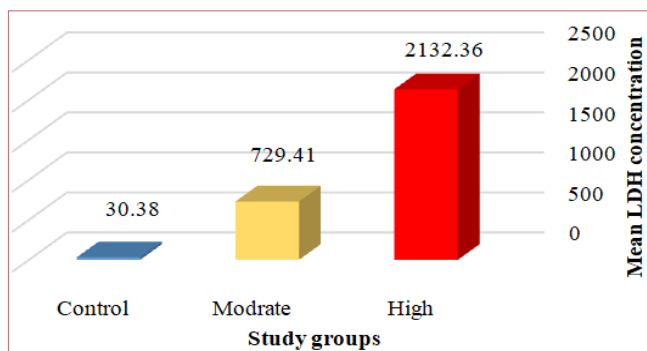
**Determination of LDH Level Concentration**

The mean differences of LDH level concentration between study groups A,B, and C were shown in Table 1 and Figure 1. The results show that there was a significant increase in LDH level concentration in group A as compared to groups B and C (P=0.001).

**Table 1. Determination of LDH Level Concentration among Study Groups**

Biomarker	Study Groups			P value
LDH U/L	Group A (25)	Group B(25)	Group C (25)	0.001*
	Mean ± SD	Mean ± SD	Mean ± SD	
	630.84 ± 212.48	254.44 ± 21.25	152.63 ± 36.97	

Normal LDH levels range from 140 U/L to 280 U/L. Group A: patients suffering from coronavirus with severe symptoms; Group B: patients suffering from coronavirus with mild symptoms; and Group C: healthy patients as control. Pvalue ≤ 0.05 was significant.



**Figure 1. Bar Diagram showing LDH Level Concentration among Study Groups**

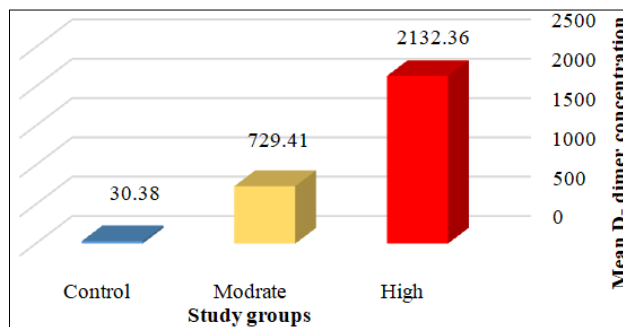
**Determination of D-dimer Level Concentration**

Table 2 and Figure 2 shows D-dimer level concentration, which is significantly higher in group A as compared to groups B and C (P=0.004).

**Table 2. Determination of D-dimer Level Concentration among Study Groups**

Biomarker	Study Groups			P value
D-dimer ng/L	Group A (25)	Group B (25)	Group C (25)	0.004*
	Mean ± SD	Mean ± SD	Mean ± SD	
	1531.57 ± 2423.89	528.91 ± 10.73	203.94 ± 66.69	

Normal D-dimer levels < 500 ng/L. Group A: patients suffering from coronavirus with severe symptoms; Group B: patients suffering from coronavirus with mild symptoms; and Group C: healthy patients as control. Pvalue ≤ 0.05 was significant.



**Figure 2. Bar diagram showing D-dimer Level Concentration among Study Groups**

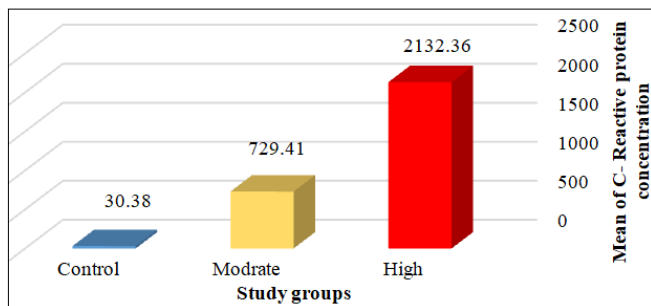
**Table 3. Determination of C-reactive Protein Level Concentration among Study Groups**

Biomarker	Study Groups			P value
CRP mg/L	Group A (25)	Group B(25)	Group C (25)	0.001*
	Mean ± SD	Mean ± SD	Mean ± SD	
	17.77 ± 4.44	2.20 ± 0.62	0.43 ± 0.28	

Normal CRP levels < 6.0 mg/L. Group A: patients suffering from coronavirus with severe symptoms; Group B: patients suffering from coronavirus with mild symptoms; and Group C: healthy patients as control. P value ≤ 0.05 was significant.

**Determination of C-reactive Protein Level Concentration**

Table 3 and Figure 3 show C-reactive protein level concentration. It shows a significant increase in C-reactive protein level concentration in group A as compared to groups B and C (P=0.001).



**Figure 3. Bar Diagram showing Comparison between C-reactive Protein Level Concentration among Study Groups**

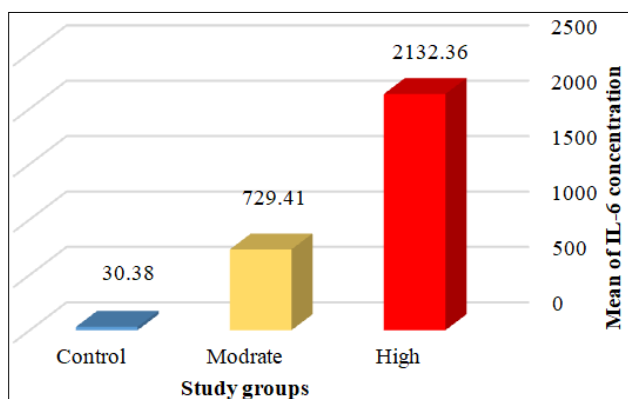
### Determination of IL-6 Level Concentration

Table 4 and Figure 4 show IL-6 level concentration among study and control groups. There is a significant increase in IL-6 level in group A compared as compared to groups B and C (P=0.001).

**Table 4. Determination of IL-6 Level Concentration among Study and Control Groups**

Biomarker	Study Groups			P value
IL-6 pg/L	Group A (25)	Group B (25)	Group C (25)	0.001*
	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	
	960.08 $\pm$ 170.24	729.41 $\pm$ 15.09	190.86 $\pm$ 79.63	

Normal IL-6 levels < 7.0 pg/L. Group A: patients suffering from coronavirus with severe symptoms; Group B: patients suffering from coronavirus with mild symptoms; and Group C: healthy patients as control. P value  $\leq$  0.05 was significant.



**Figure 4. Bar Diagram showing Comparison of IL-6 Level Concentration among Study and Control Groups**

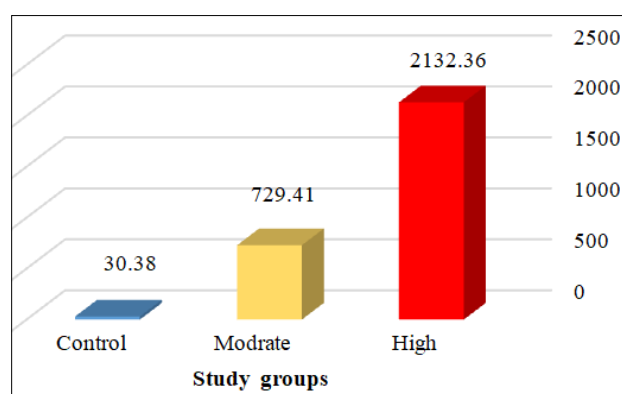
### Determination of Ferritin Level Concentration

Table 5 and Figure 5 show ferritin level concentration among study and control groups. There is a significant increase in ferritin level concentration in group A as compared to groups B and C (P=0.001).

**Table 5. Determination of Ferritin Level Concentration among Study and Control Groups**

Biomarker	Study Groups			P value
Ferritin ng/L	Group A (25)	Group B (25)	Group C (25)	0.001*
	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	
	2132.36 $\pm$ 1309.35	729.41 $\pm$ 15.09	30.38 $\pm$ 11.03	

Normal male ferritin levels range from 25-350 ng/L. Group A: patients suffering from coronavirus with severe symptoms; Group B: patients suffering from coronavirus with mild symptoms; and Group C: healthy patients as control. P value  $\leq$  0.05 was significant.



**Figure 5. Bar Diagram showing Comparison of Ferritin Level Concentration among Study Control Groups**

### Discussion

When the virus enters the body, it wants a special receiver to support it into the cell, which is called the vasoconstrictive changes in the enzymes, ACE 2.<sup>18</sup> ACE 2 is a gift within the respiratory organ on the surface of the alveolar cells.<sup>19</sup> They need three sorts of alveolar cells: Type 1: accountable for gas exchange with blood vessels (gas exchange), Type 2: it's responsible for the assembly of surfactant, that may be a mixture of proteins and fats that cut back the physical phenomenon of the air sac and kind 3: immune cells, which are macrophages.<sup>20</sup> The coronavirus envelope contains proteins referred to as spikes and specials (S\_Spike) that facilitate the virus bind to ACE2.<sup>21</sup> The virus' genetic material enters the cell's interior and also the cell is controlled to provide the virus' proteins, and so the virus multiplies and the cell dies.<sup>22</sup> After two alveolar cells have passed away, they throw out certain inflammatory mediators for alleged substances. These substances stimulate the dominant "macrophages" of the immune cells, and they secrete three immune substances called "cytokine" after stimulating the macrophages: interleukin 1, interleukin 6 and the mortification problem with the neoplasm.<sup>23</sup> The symptoms associated with Coronavirus infection are caused by the 3 substances when they reach the bloodstream.<sup>24</sup> The blood

vessels also enlarge the cyst as permeability, vasodilation and capillary permeability of these vessels are increased exaggeratedly.<sup>25</sup> This leads to “alveolar oedema” leading to drive and breathlessness. Loss of a chemical substance leads to an increase of the vesicle’s physicality and thus to the collapse of the Alveolar system. It also causes breathlessness and hypoxia.<sup>26</sup> In addition, the flow of “neutrophils” into the infectious agent infection location as an associate degree reaction to these three substances increases.<sup>27</sup> Cells of neutrophils kill viruses with the secretion of two substances like ROS, Proteases. However, in addition, the second article breaks down a number of alveolar cells responsible for exchanges of gas and causes a condition called “cough” consolidation.<sup>28</sup> The 3 substances, “IL-1, IL-6, and TNF” attend the “hypothalamus” and so increase the vital sign and cause “fever” symptoms. In extreme cases, the “SIRS” can occur. Syndrome of generalized inflammatory reaction. 29 This results in septic shock. Syndrome with multiple organ disease. Particularly the excretory organ, which results in renal failure.<sup>30</sup> One of these biomarkers of concern is feed dehydrogenase (LDH), particularly as higher LDH levels have been linked to worse outcomes among patients with previously different infective agent infections.<sup>31</sup> Early knowledge of COVID-19 patients led to significant differences in LDH levels, while not severe sickness, among patients with associated degrees.<sup>32</sup> A group analysis of the literature revealed was therefore conducted in order to investigate the possible connection between exaggerated LDH values for patients with COVID-19.<sup>33</sup> In severe, non-severe cases of COVID-19 many studies compared elevated LDH values.<sup>34,35</sup> In almost all organs, LDH is a living catalyst found in cellular components that catalyses pyruvate and lactate interconversion with concurrent NADH and NAD<sup>+</sup> interconversion.<sup>36</sup> The catalyst is comprised of 2 major sub-units (i.e., A and B) and is donated to humans in 5 separate isozymes (CARL-1, RR-2-2, LRH-3, RCC LDH-3, RCL and PRH-2 and LDH-5).<sup>37</sup> The catalyst is used in the human body in the cardiomyocytes and in the panoramic system. While LDH has been used historically since the 1960s as a marker of viscus injuries, abnormal values can come out of multiple organ wounds and reduced action with glycolytic upregulation.<sup>38</sup> The acidic extracellular pH caused by infection feed and tissue damage triggers metalloprotease activation and increases the medium phagocyte ontogeny.<sup>39</sup> But the contribution to the LDH elevation determined in COVID-19 by the different LDH isoenzymes was not determined. LDH levels in nephrosis and cardiac muscle injury are also increased in thrombotic microangiopathy.<sup>40</sup> In addition, high dimer concentrations and blood disorder have been reported in serious COVID-19 patients, indicating that hyper coagulating conditions may also contribute to the severity of illness and deaths.<sup>41</sup> COVID-19 patients had main fever clinical

symptoms, most of which were sensitive and many severe cases. Due to the variability of symptoms and imaging findings, and the variable level of sickness progression, the condition and prognosis for COVID-19 patients were sophisticated.<sup>42</sup> The dynamic changes of the peripheral blood clotting indices in patients who loved D-dimer were observed in COVID-19, which showed that level D-dimers could well be used to forecast COVID-19 severity and prognosis.<sup>43</sup> D-dimer is that fibrinolytic degradation product and high levels indicate that the body is hyper coagulated and the secondary decay is very useful in determining thrombosis. It has been reported that COVID-19 patients have a hypercoagulable condition.<sup>44</sup> Further, in patients with severe COVID-19 the incidence of blood vessel occlusion (VTE) was 25%, while embolism was diagnosed in 30% of COVID-19 patients.<sup>45</sup> Additional levels of D-dimer were exaggerated within the blood of COVID-19 patients with CVA.<sup>46</sup> The next D-dimer value also indicated that the patient’s condition could be very serious, and even combined with various complications.<sup>47</sup> For COVID-19 patients, there are attainable reasons for increasing dimer values: The infection causes pro-inflammatory cytokines to be released so that an inflammatory storm is infected.<sup>48</sup> In plasma, love IL-2, IL-6, G-CSF, IP-10, MCP-1, MIP-1A and TNF- $\alpha$  were higher especially in severe COVID-19 patients, with apace and extremely active T cells, macrophages and natural cells, with overruns in the immunodeficiency cell or non-immune and with more than a hundred and fifty infectious cytokines and Checells and with over 100 inflammatory cells. In some COVID-19 patients, hypoxia is completely different, and inflammation leads to occlusion or over-consumption of gas.<sup>50</sup> The absolute demand for oxygen will increase in the whole of the abnormal hemodynamics which triggers and ends up in thrombosis, both molecular and cellular. Severe infection or acute septic inflammation could also influence coagulation of the blood, love excessive levels of proteolytic enzyme (PAI-1), and excessive tenured dissolution.<sup>52</sup> Serum globulin may be an inflammatory substance produced by the liver. The C-reactive protein examines how much this protein is measured in the blood. The test can help diagnose acute and chronic inflammatory conditions.<sup>53</sup> In COVID-19, the handwriting misunderstands the importance of high IL-6. Although IL-6 is generally greater in various inflammatory conditions, the pathophysiology of COVID-19 is restricted in this finding. Different studies have demonstrated the association between initial IL-6 levels greater than 80 pg/mL and the results of love metastasis failure and death.<sup>54</sup> The authors fail to discuss prognostic models such as ISARIC-4C or associated hyperinflammatory syndrome COVID-19, which underline the prognostic marks of sugar globulin and IL-6.<sup>55</sup> Significantly high levels of IL-6 body fluid do not appear to be the clinical reaction requirement for IL-6

blockage. Median IL-6 pretreatment concentrations are only moderately elevated in approved sickness indications such as multicenter Castleman's disease and large cell arteritis.<sup>56</sup> While the results of the IL-6 blockade irregularly controlled studies have been largely negative, various immunomodulatory therapy studies investigating each short-term outcome are being conducted.<sup>57</sup> COVID-19 is a heterogenic disease that can cause many manifestations of immune disorder in children, beginning with the protein storm, via alleged long-haul syndrome and inflammatory multisystem disorder.<sup>58</sup> Body fluid IL-6 remains the most effective COVID-19 severity biomarker on the market and still has good guiding potential for this disease. The COVID-19 fatal effects during cytokine storm syndrome have been reported to depend on the severity of the disease. A body fluid protein content assessment is carried out by this cytokine storm.<sup>59</sup> A load of infectious agents or microorganisms in the body indicates increased amounts of ferritin or hyperferritinemia. A condition activating macrophages to secrete proteins can be hyperferritinemia or hyperferritinemic syndrome, which may be a sign of serious sickness, and in severe cases, may cause a cytokine storm.<sup>60</sup> During cytokine storm syndrome the fatal results of COVID-19 are present. The severity of the disease depending on cytokine tempest syndrome has thus been reported. A complete evaluation of serum ferritin levels can be carried out in this cytokine storm.<sup>61</sup> The gravity of COVID-19 might best be determined by serum ferritin levels. Many people suffering from polygenic disorder have high levels of body fluid proteins, which are likely to have serious COVID-19 complications.<sup>62</sup> The median values of serum ferritin from several recent studies have exceeded COVID-19 detection limits during the entire hospital period, indicating that the levels of ferritin in all hospitals have been continuously exaggerated.<sup>63</sup>

## Conclusion

Increased LDH values were linked to an increase in COVID-19 toxicity. On the basis of D-dimer, the probability of mortality can be determined. C-responsive protein and ferritin serum exercises were significantly increased in COVID-19 patients compared to those with mild side effects of COVID. IL-6 is a key immunomodulatory cytokine in both sound and infected tissues.

**Source of Funding:** None

**Conflict of Interest:** None

## References

1. Pityana NB. A theological statement on the coronavirus pandemic: living the faith responsibly. *Relig Theol.* 2020;27(3-4):329-58. [Google Scholar]
2. Ambrose PM. High throughput genetics and characterization of an RNA Arbovirus, Sindbis virus, using accurate next-generation sequencing of viral evolution and RNA enrichment [dissertation]. Weill Medical College of Cornell University. ProQuest Dissertations Publishing; 2020. [Google Scholar]
3. Rangayasami A, Kannan K, Murugesan S, Radhika D, Sadasivuni KK, Reddy KR, Raghu AV. Influence of nanotechnology to combat against COVID-19 for global health emergency: a review. *Sens Int.* 2021;2:100079. [PubMed] [Google Scholar]
4. Ortiz-Prado E, Simbaña-Rivera K, Gómez-Barreno L, Rubio-Neira M, Guaman LP, Kyriakidis NC, Muslin C, Jaramillo AMG, Barba-Ostria C, Cevallos-Robalino D, Sanches-SanMiguel H, Unigarro L, Zalakeviciute R, Gadian N, López-Cortés A. Clinical, molecular and epidemiological characterization of the SARS-CoV2 virus and the Coronavirus disease 2019 (COVID-19), a comprehensive literature review. *Diagn Microbiol Infect Dis.* 2020 Sep;98(1):115094. [PubMed] [Google Scholar]
5. Qasim QA, Hussein HH, Shari FH, Al-Salman HNK, Ahmed, GS, Jeber MA. General study: the effect of corpulence and persistent sicknesses on the seriousness of the diseases with COVID-19. *Int J Pharm Res.* 2020;12(3):3539-64. [Google Scholar]
6. Ali SR, Dobbs TD, Whitaker IS. Webinars in plastic and reconstructive surgery training-a review of the current landscape during the COVID-19 pandemic. *J Plast Reconstr Aesthet Surg.* 2020 Jul;73(7):1357-404. [PubMed] [Google Scholar]
7. Bae S, Kim SR, Kim MN, Shim WJ, Park SM. Impact of cardiovascular disease and risk factors on fatal outcomes in patients with COVID-19 according to age: a systematic review and meta-analysis. *Heart.* 2021 Mar;107(5):373-80. [PubMed] [Google Scholar]
8. Guery B, Poissy J, el Mansouf L, Séjourné C, Ettahar N, Lemaire X, Vuotto F, Goffard A, Behillil S, Enouf V, Caro V, Mailles A, Che D, Manuguerra JC, Mathieu D, Fontanet A, van der Werf S; MERS-CoV study group. Clinical features and viral diagnosis of two cases of infection with Middle East Respiratory Syndrome coronavirus: a report of nosocomial transmission. *Lancet.* 2013 Jun;381(9885):2265-72. [PubMed] [Google Scholar]
9. Huang J, Mao T, Li S, Wu L, Xu X, Li H, Xu C, Su F, Dai J, Shi J, Cai J. Long period dynamics of viral load and antibodies for SARS-CoV-2 infection: an observational cohort study. *MedRxiv.* 2020 Jan 1. [Google Scholar]
10. Assadullah Z, Kumar A, Kumar P, Barik M. Guidelines and protocol development for Molecular Targeted Therapy (MMT) for COVID-19 patients. [Google Scholar]
11. Zuo YY, Uspal WE, Wei T. Airborne transmission of COVID-19: aerosol dispersion, lung deposition, and virus-receptor interactions. *ACS Nano.* 2020 Nov;acs.nano.0c08484. [PubMed] [Google Scholar]
12. Schwartz DA. An analysis of 38 pregnant women with

- COVID-19, their newborn infants, and maternal-fetal transmission of SARS-CoV-2: maternal coronavirus infections and pregnancy outcomes. *Arch Pathol Lab Med*. 2020 Jul;144(7):799-805. [PubMed] [Google Scholar]
13. Zhang W, Zhao Y, Zhang F, Wang Q, Li T, Liu Z, Wang J, Qin Y, Zhang X, Yan X, Zeng X, Zhang S. The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): the perspectives of clinical immunologists from China. *Clin Immunol*. 2020 May;214:108393. [PubMed] [Google Scholar]
  14. Menter T, Haslbauer JD, Nienhold R, Savic S, Hopfer H, Deigendesch N, Frank S, Turek D, Willi N, Pargger H, Bassetti S, Leuppi JD, Cathomas G, Tolnay M, Mertz KD, Tzankov A. Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction. *Histopathology*. 2020 Aug;77(2):198-209. [PubMed] [Google Scholar]
  15. Xu K, Cui K, Young LH, Wang YF, Hsieh YK, Wan S, Zhang J. Air quality index, indicator air pollutants and impact of COVID-19 event on the air quality near central China. *Aerosol Air Qual Res*. 2020;20(6):1204-21. [Google Scholar]
  16. Bai HX, Wang R, Xiong Z, Hsieh B, Chang K, Halsey K, Tran TML, Choi JW, Wang DC, Shi LB, Mei J, Jiang XL, Pan J, Zeng QH, Hu PF, Li YH, Fu FX, Huang RY, Sebro R, Yu QZ, Atalay MK, Liao WH. Artificial intelligence augmentation of radiologist performance in distinguishing COVID-19 from pneumonia of other origin at chest CT. *Radiology*. 2020 Sep;296(3):E156-65. [PubMed] [Google Scholar]
  17. Upadhyay M, Yadav S, Nagaraj K, Patil S. Treatment effects of mini-implants for en-masse retraction of anterior teeth in bialveolar dental protrusion patients: a randomized controlled trial. *Am J Orthod Dentofacial Orthop*. 2008 Jul;134(1):18-29.e1. [PubMed] [Google Scholar]
  18. Krishna Kumar B, Rana S. COVID 19 in INDIA: strategies to combat from combination threat of life and livelihood. *J Microbiol Immunol Infect*. 2020 Jun;53(3):389-91. [PubMed] [Google Scholar]
  19. Pfefferbaum B, North CS. Mental health and the COVID-19 pandemic. *N Engl J Med*. 2020;383:510-2. [Google Scholar]
  20. Hsu LY, Chia PY, Lim JF. The Novel corona virus (SARS-CoV-2) epidemic. *Ann Acad Med Singap*. 2020 Mar;49(3):105-7. [PubMed] [Google Scholar]
  21. Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, Daszak P. Global Trends in emerging infectious diseases. *Nature*. 2008 Feb;451(7181):990-3. [PubMed] [Google Scholar]
  22. Lupia T, Scabini S, Mornese Pinna S, Di Perri G, De Rosa FG, Corcione S. 2019 novel corona virus (2019-nCoV) outbreak: a new challenge. *J Glob Antimicrob Resist*. 2020 Jun;21:22-7. [PubMed] [Google Scholar]
  23. Madhav N, Oppenheim B, Gallivan M, Mulembakani P, Rubin E, Wolfe N, Jamison DT, Gelband H, Horton S, Jha P, Laxminarayan R, Mock CN, Nugent R, editors. *Pandemics: risks, impacts, and mitigation*. In: *Disease control priorities: improving health and reducing poverty*. 3rd ed. Washington (DC): The International Bank for Reconstruction and Development, The World Bank; 2017. [PubMed] [Google Scholar]
  24. Omer SB, Malani P, del Rio C. The COVID-19 pandemic in the US: a clinical update. *JAMA*. 2020 May;323(18):1767-8. [PubMed] [Google Scholar]
  25. De P. COVID-19, new normal and India [Internet]. *The Economic Times*; 2020 [cited 2020 Apr 9]. Available from: <https://economictimes.indiatimes.com/blogs/et-commentary/covid-19-new-normal-and-india/>
  26. Wang H, Wang S, Yu K. COVID-19 infection epidemic: the medical management strategies in Heilongjiang Province, China. *Crit Care*. 2020 Mar;24(1):107. [PubMed] [Google Scholar]
  27. Yang Y, Bazhin AV, Werner J, Karakhanova S. Reactive oxygen species in the immune system. *Int Rev Immunol*. 2013 Jun;32(3):249-70. [PubMed] [Google Scholar]
  28. Smith JA. Neutrophils, host defense, and inflammation: a double-edged sword. *J Leukoc Biol*. 1994 Dec;56(6):672-86. [PubMed] [Google Scholar]
  29. Bryan N, Ahswain H, Smart N, Bayon Y, Wohlert S, Hunt JA. Reactive oxygen species (ROS) - a family of fate deciding molecules pivotal in constructive inflammation and wound healing. *Eur Cell Mater*. 2012 Sep;24:249-65. [PubMed] [Google Scholar]
  30. Henry BM, Aggarwal G, Wong J, Benoit S, Vikse J, Plebani M, Lippi G. Lactate dehydrogenase levels predict coronavirus disease 2019 (COVID-19) severity and mortality: a pooled analysis. *Am J Emerg Med*. 2020 Sep;38(9):1722-6. [PubMed] [Google Scholar]
  31. Malik P, Patel U, Mehta D, Patel N, Kelkar R, Akrmah M, Gabrilove JL, Sacks H. Biomarkers and outcomes of COVID-19 hospitalisations: systematic review and meta-analysis. *BMJ Evid Based Med*. 2021 Jun;26(3):107-8. [PubMed] [Google Scholar]
  32. Terpos E, Ntanasis-Stathopoulos I, Elalamy I, Kastritis E, Sergentanis TN, Politou M, Psaltopoulou T, Gerotziapas G, Dimopoulos MA. Hematological findings and complications of COVID-19. *Am J Hematol*. 2020 Jul;95(7):834-47. [PubMed] [Google Scholar]
  33. Sun J, Su J, Xie Y, Yin MT, Huang Y, Xu L, Zhou Q, Zhu B. Plasma IL-6/IL-10 ratio and IL-8, LDH, and HBDH level predict the severity and the risk of death in AIDS patients with pneumocystis pneumonia. *J Immunol*

- Res. 2016;2016:1583951. [PubMed] [Google Scholar]
34. Wei XL, Zhang DS, He MM, Jin Y, Wang DS, Zhou YX, Bai L, Li ZZ, Luo HY, Wang FH, Xu RH. The predictive value of alkaline phosphatase and lactate dehydrogenase for overall survival in patients with esophageal squamous cell carcinoma. *Tumour Biol.* 2016 Feb;37(2):1879-87. [PubMed] [Google Scholar]
  35. Mishra D, Banerjee D. Lactate dehydrogenases as metabolic links between tumor and stroma in the tumor microenvironment. *Cancers (Basel).* 2019 May;11(6):750. [PubMed] [Google Scholar]
  36. Ding J, Karp JE, Emadi A. Elevated lactate dehydrogenase (LDH) can be a marker of immune suppression in cancer: interplay between hematologic and solid neoplastic clones and their microenvironments. *Cancer Biomark.* 2017 Jul;19(4):353-63. [PubMed] [Google Scholar]
  37. Akagawa M, Minematsu K, Shibata T, Kondo T, Ishii T, Uchida K. Identification of lactate dehydrogenase as a mammalian pyrroloquinolinequinone (PQQ)-binding protein. *Sci Rep.* 2016 May;6:26723. [PubMed] [Google Scholar]
  38. Baumgart E, Fahimi HD, Stich A, Völkl A. L-lactate dehydrogenase A4-and A3B isoforms are bona fide peroxisomal enzymes in rat liver: evidence for involvement in intraperoxisomal NADH reoxidation. *J Biol Chem.* 1996 Feb;271(7):3846-55. [PubMed] [Google Scholar]
  39. Maldonado V, Hernandez-Ramírez C, Oliva-Pérez EA, Sánchez-Martínez CO, Pimentel-González JF, Molina-Sánchez JR, Jiménez-Villalba YZ, Chávez-Alderete J, Loza-Mejía MA. Pentoxifylline decreases serum LDH levels and increases lymphocyte count in COVID-19 patients: results from an external pilot study. *Int Immunopharmacol.* 2021 Jan;90:107209. [PubMed] [Google Scholar]
  40. Han Y, Zhang H, Mu S, Wei W, Jin C, Tong C, Song Z, Zha Y, Xue Y, Gu G. Lactate dehydrogenase, an independent risk factor of severe COVID-19 patients: a retrospective and observational study. *Aging (Albany NY).* 2020 Jun;12(12):11245-58. [PubMed] [Google Scholar]
  41. Klein R, Nagy O, Tóthová C, Chovanová F. Clinical and diagnostic significance of lactate dehydrogenase and its isoenzymes in animals. *Vet Med Int.* 2020 Jun;2020:5346483. [PubMed] [Google Scholar]
  42. Fu J, Kong J, Wang W, Wu M, Yao L, Wang Z, Jin J, Wu D, Yu X. The clinical implication of dynamic neutrophil to lymphocyte ratio and D-dimer in COVID-19: a retrospective study in Suzhou China. *Thromb Res.* 2020 Aug;192:3-8. [PubMed] [Google Scholar]
  43. Ye W, Chen G, Li X, Lan X, Ji C, Hou M, Zhang D, Zeng G, Wang Y, Xu C, Lu W, Cui R, Cai Y, Huang H, Yang L. Dynamic changes of D-dimer and neutrophil-lymphocyte count ratio as prognostic biomarkers in COVID-19. *Respir Res.* 2020 Jul;21(1):169. [PubMed] [Google Scholar]
  44. He X, Yao F, Chen J, Wang Y, Fang X, Lin X, Long H, Wang Q, Wu Q. The poor prognosis and influencing factors of high D-dimer levels for COVID-19 patients. *Sci Rep.* 2021 Jan;11(1):1830. [PubMed] [Google Scholar]
  45. Bao J, Li C, Zhang K, Kang H, Chen W, Gu B. Comparative analysis of laboratory indexes of severe and non-severe patients infected with COVID-19. *Clin Chim Acta.* 2020 Oct;509:180-94. [PubMed] [Google Scholar]
  46. Cerdà P, Ribas J, Iriarte A, Mora-Luján JM, Torres R, Del Río B, Jofre HI, Ruiz Y, Huguet M, Fuset MP, Martínez-Yélamos S, Santos S, Llecha N, Corbella X, Riera-Mestre A. Blood test dynamics in hospitalized COVID-19 patients: potential utility of D-dimer for pulmonary embolism diagnosis. *PLoS One.* 2020 Dec;15(12):e0243533. [PubMed] [Google Scholar]
  47. Soy M, Keser G, Atagündüz P, Tabak F, Atagündüz I, Kayhan S. Cytokine storm in COVID-19: pathogenesis and overview of anti-inflammatory agents used in treatment. *Clin Rheumatol.* 2020 Jul;39(7):2085-94. [PubMed] [Google Scholar]
  48. Tan M, Liu Y, Zhou R, Deng X, Li F, Liang K, Shi Y. Immunopathological characteristics of coronavirus disease 2019 cases in Guangzhou, China. *Immunology.* 2020 Jul;160(3):261-8. [PubMed] [Google Scholar]
  49. Herrmann J, Mori V, Bates JHT, Suki B. Modeling lung perfusion abnormalities to explain early COVID-19 hypoxemia. *Nat Commun.* 2020 Sep;11(1):4883. [PubMed] [Google Scholar]
  50. Moriarty PM, Gorby LK, Stroes ES, Kastelein JP, Davidson M, Tsimikas S. Lipoprotein (a) and its potential association with thrombosis and inflammation in COVID-19: a testable hypothesis. *Curr Atheroscler Rep.* 2020 Jul;22(9):48. [PubMed] [Google Scholar]
  51. Du Clos TW. Function of C-reactive protein. *Ann Med.* 2000 May;32(4):274-8. [PubMed] [Google Scholar]
  52. Channappanavar R, Perlman S. Age-related susceptibility to Coronavirus infections: role of impaired and dysregulated host immunity. *J Clin Invest.* 2020 Dec;130(12):6204-13. [PubMed] [Google Scholar]
  53. Szabo PA, Dogra P, Gray JI, Wells SB, Connors TJ, Weisberg SP, Krupska I, Matsumoto R, Poon MML, Idzikowski E, Morris SE, Pasin C, Yates AJ, Ku A, Chait M, Davis-Porada J, Guo XV, Zhou J, Steinle M, Mackay S, Saqi A, Baldwin MR, Sims PA, Farber DL. Longitudinal profiling of respiratory and systemic immune responses reveals myeloid cell-driven lung inflammation in severe COVID-19. *Immunity.* 2021 Apr;54(4):797-814.e6. [PubMed] [Google Scholar]
  54. Coperchini F, Chiovato L, Ricci G, Croce L, Magri F, Rotondi M. The Cytokine storm in COVID-19: further advances in our understanding the role of specific



- chemokines involved. *Cytokine Growth Factor Rev.* 2021 Apr;58:82-91. [PubMed] [Google Scholar]
55. Rendeiro AF, Casano J, Vorkas CK, Singh H, Morales A, DeSimone RA, Ellsworth GB, Soave R, Kapadia SN, Saito K, Brown CD, Hsu J, Kyriakides C, Chiu S, Cappelli LV, Cacciapuoti MT, Tam W, Galluzzi L, Simonson PD, Elemento O, Salvatore M, Inghirami G. Profiling of immune dysfunction in COVID-19 patients allows early prediction of disease progression. *Life Sci Alliance.* 2020 Dec;4(2):e202000955. [PubMed] [Google Scholar]
56. Fraga-Silva TFC, Maruyama SR, Sorgi CA, Russo EMS, Fernandes APM, de Barros Cardoso CR, Faccioli LH, Dias-Baruffi M, Bonato VLD. COVID-19: integrating the complexity of systemic and pulmonary immunopathology to identify biomarkers for different outcomes. *Front Immunol.* 2021 Jan;11:599736. [PubMed] [Google Scholar]
57. Taefehshokr N, Taefehshokr S, Heit B. Mechanisms of dysregulated humoral and cellular immunity by SARS-CoV-2. *Pathogens.* 2020 Dec;9(12):1027. [PubMed] [Google Scholar]
58. Castiglione F, Deb D, Srivastava AP, Liò P, Liso A. From infection to immunity: understanding the response to SARS-CoV2 through in-silico modeling. *Front Immunol.* 2021 Sep;12:646972. [PubMed] [Google Scholar]
59. To KK, Sridhar S, Chiu KH, Hung DL, Li X, Hung IF, Tam AR, Chung TW, Chan JF, Zhang AJ, Cheng VC, Yuen KY. Lessons learned one year after SARS-CoV-2 emergence leading to COVID-19 pandemic. *Emerg Microbes Infect.* 2021 Dec;10(1):507-35. [PubMed] [Google Scholar]
60. Brunet-Ratnasingham E, Anand SP, Gantner P, Dyachenko A, Moquin-Beaudry G, Brassard N, Beaudoin-Bussièrès G, Pagliuzza A, Gasser R, Benlarbi M, Point F, Prévost J, Laumaea A, Niessl J, Nayrac M, Sannier G, Orban C, Messier-Peet M, Butler-Laporte G, Morrison DR, Zhou S, Nakanishi T, Boutin M, Descôteaux-Dinelle J, Gendron-Lepage G, Goyette G, Bourassa C, Medjahed H, Laurent L, Rébillard RM, Richard J, Dubé M, Fromentin R, Arbour N, Prat A, Larochelle C, Durand M, Richards JB, Chassé M, Tétreault M, Chomont N, Finzi A, Kaufmann DE. Integrated immunovirological profiling validates plasma SARS-CoV-2 RNA as an early predictor of COVID-19 mortality. *Sci Adv.* 2021 Nov;7(48):eabj5629. [PubMed] [Google Scholar]
61. Heinz C, Miesbach W, Herrmann E, Sonntagbauer M, Raimann FJ, Zacharowski K, Weber CF, Adam EH. Greater fibrinolysis resistance but no greater platelet aggregation in critically ill COVID-19 patients. *Anesthesiology.* 2021 Mar;134(3):457-67. [PubMed] [Google Scholar]