

Review Article

# Hypoxia and Cigarette Smoke - A Combined Threat to the Brain: Role of Toll-like Receptor 4 in Neuroinflammatory Changes

Jayalakshmi Krishnan<sup>1</sup>, SK Farhat<sup>2</sup>, Rajalakshmi Anbalagan<sup>3</sup>

<sup>1</sup>Assistant Professor, <sup>2,3</sup>Department of Life Sciences, Central University of Tamil Nadu, Thiruvavur, India.

DOI: <https://doi.org/10.24321/0019.5138.202291>

## I N F O

### Corresponding Author:

Jayalakshmi Krishnan, Department of Life Sciences, Central University of Tamil Nadu, Thiruvavur, India.

### E-mail Id:

[jayalakshmi@cutn.ac.in](mailto:jayalakshmi@cutn.ac.in)

### Orcid Id:

<https://orcid.org/0000-0003-3098-5755>

### How to cite this article:

Krishnan J, Farhat SK, Anbalagan R. Hypoxia and Cigarette Smoke - A Combined Threat to the Brain: Role of Toll-like Receptor 4 in Neuroinflammatory Changes. J Commun Dis. 2022;54(3):67-74.

Date of Submission: 2022-07-20

Date of Acceptance: 2022-09-23

## A B S T R A C T

The mammalian brain is composed of 20% of oxygen. The depletion in the level of oxygen may lead to hypoxia, resulting in neurological changes. Recent findings of Toll-like receptors (TLR) such as TLR-2, 3, 4 and 8 in the mammalian nervous system showed neuroinflammation in the brain. The activation of TLR4 by both exogenous and endogenous ligands leads to various inflammatory diseases like atherosclerosis, Crohn's disease, ulcerative colitis, rheumatoid arthritis and prostate cancer. This study confirms the involvement of NF- $\kappa$ B in TLR4-axis-induced neuroinflammation in the brain. Cigarette Smoke contains many toxic chemicals such as carcinogenic compounds which are injurious to health, causing death worldwide. The role of TLR4 in activating the Brain during Cigarette Smoke exposure is not yet understood. Hence this study reviewed the use of the cell culture in vitro model as well as in vivo mouse model of TLR4-/- mouse exposed to hypobaric hypoxia to test hypothesis.

**Methodology:** This review explored articles from the past 15 years to study the action of toll-like receptor 4 in neuroinflammatory changes. Those articles which lacked full texts were excluded from the review. A total of 61 articles were enrolled in this review article.

**Results:** The recent findings of TLR in Neurons, showed the involvement of external stimuli such as Cigarette smoke causing various neurological disorders and brain inflammation. The role of TLR4 in activating the brain during acute and chronic cigarette smoke exposure is not yet investigated and also the neuroimmune mechanisms of cigarette smoke are not yet clearly understood.

**Conclusion:** Smoking-related neurological disorders require a proper understanding of the mechanisms of such damage for therapeutic development. Hence research should be carried out to study the TLR4 mechanisms and pathways in various inflammatory diseases.

**Keywords:** TLR4, NF-Kb, (IFN- $\beta$ ), Janus Associated Kinases/ Signal Transducer and Activator of Transcription (JAK/STAT), Pathogen Associated Molecular Patterns (PAMPS), Cigarette Smoke, Neuroinflammation

## Introduction

Hypobaric hypoxia is injurious to health. The brain is highly susceptible to hypoxia; it is only 2% of body weight and consumes 20% of oxygen. Hypoxia causes memory impairment leading to learning deficiencies, anterograde amnesia<sup>1</sup> and cognitive functions in humans. Hypobaric hypoxic effects in the brain are characterized by changes in the biochemical components, apoptosis, neurotransmitter release and neuroinflammation.<sup>2</sup> Brain inflammation is a consequence of hypoxia. The causes and mechanisms of brain inflammation following hypoxia are not well understood. Hypoxia stress itself might act as neuroinflammation. Toll-like receptors (TLRs) are archetypal pattern recognition receptors expressed in the immune system and regulate thousands of genes that are involved in innate and adaptive immune responses to fight against microbial infection.<sup>3</sup> TLRs are not only expressed in the immune system, the non-immune cells such as brain cells, endothelial cells and fibroblasts, hepatocytes also express them.<sup>4</sup> Astrocytes express TLR4 and it has been investigated for pro-inflammatory mediators.<sup>5</sup> Microglia expresses all known TLRs and experiments indicate that they can robustly respond to both TLR4 and TLR3 ligands.<sup>6</sup> Recently some studies pointed out that neurons, as well as neural progenitor cells, can also express TLRs.<sup>7</sup> Further reports also reveal that TLR2, 3, 4 and 8 are expressed in neurons.<sup>8</sup> Direct stimulation of TLR4 by LPS has been reported to cause memory impairment.<sup>9</sup> Another study of hypobaric hypoxia indicated memory impairment in rats.<sup>10</sup> However, the cause of this is not clearly understood. The studies<sup>11,12</sup> using single-Ig-IL-1 related receptor (SIGIRR) KO in mice identified that overstimulation by TLR4 by High Mobility Group Box1 (HMGB1) was involved in memory dysfunctions. It is possible that increased HMGB1 levels may be involved in high altitude-induced memory impairment, by stimulating TLR4 signalling. Hence in this review, focus is on using the cell culture in vitro model as well as in vivo mouse model of TLR4-/- mouse exposed to hypobaric hypoxia to test our hypothesis.

The actual mechanisms of memory impairment was explored using the cell lines such as BV2 cells, HT22 cell line and IMA 2.1 as potential model systems to study microglial, neuronal and astrocyte functions. Thus, the study aimed to understand the non-additive responses to identify/ delineate the signalling mechanisms.

## The Pathways

The pathways are numerous in a cell; especially in CNS, the cells do interact extensively with each other by releasing the neurotransmitters, up taking the neurotransmitters, etc. A study stated that if two separate pathways are triggered by two distinct ligands, then they regulate the transcription of the same gene and the associated

stimulation with both ligands must result in an additive response which should correspond to the sum of the separate responses.<sup>13</sup> Despite, that, if the pathways intersect or modulate each other, then the concomitant addition of ligands will yield a non-additive response which will be either greater or less than the expected additive response. For example, the signalling pathways used by exogenous and endogenous ligands of KDO can be modulated by the presence of interferon- $\beta$  (IFN- $\beta$ ) and/or 8-bromoadenosine-3',5'-cyclic monophosphate (8-Br). IFN- $\beta$  signalling occurs via Janus Associated Kinases/ Signal Transducer and Activator of Transcription (JAK/STAT) pathways in neurons during hypoxia<sup>14</sup> and during oxygen glucose deprivation. 8-Br increases cAMP levels in the neurons.<sup>15</sup> IFN- $\beta$  and 8-Br might modulate the signalling mechanisms of TLR4 ligands in the proposed cell models. With this rationale, this present investigation was used to understand the pathway interactions and modulations among the TLR4 ligand exogenous and endogenous (such as KDO and HMGB1) individually and non-TLR4 ligand stimulation (such as IFN- $\beta$  and 8-Br in combination with KDO and HMGB1 as a double and triple combination) in neurons, astrocytes and microglia. The investigation also aimed to analyze whether pathways triggered by these ligands will be different from each other by using systems biological approaches. The present study aimed to understand the memory impairment caused by hypobaric hypoxia using KDO and HMGB1 as TLR4 ligands in vitro cell culture models.

## Methodology

This review focuses on the studies carried out across the globe. In this search, all the research articles related to toll-like receptor 4 in neuroinflammatory changes and hypoxia were explored from national and international journals. The articles were assimilated from Google scholar and PubMed using keywords such as TLR4, Nuclear Factor Kappa B, Hypoxia, Neuroinflammation, Cigarette Smoke and IFN.

This review explored articles from the past 15 years to study the action of toll-like receptor 4 in neuroinflammatory changes. Those articles which lacked full text were excluded from the review. A total of 61 articles were enrolled in this review article.

## Results

### CNS and Immune Functions

Central Nervous System (CNS) is under the monitoring of the immune cells constantly from the layer of meninges itself.<sup>24</sup> The mechanisms of those immune monitoring under various stress and disease conditions of the CNS are very poorly researched. Recently Louveuet al., (2015)<sup>24</sup> reported the presence of lymphatic vessels in the central nervous system in the dural sinuses of the mouse brain. Such discovery is a break through in the field of neuroimmunology to find new

therapeutic approaches for immune-related neurological diseases. While discussing about the trafficking of immune cells to the brain various routes of entry are emphasized. T cells and monocytes continuously influx into the brain via various routes, as the presence of Blood Brain Barrier (BBB) is not a barrier to their entry because these cells enter via post-capillary venules<sup>24</sup> thus making the brain highly vulnerable to immunological attack. Understanding such functions of CNS thus becomes a priority as it can lead to inflammation and also neurological disturbances.

### Toll-Like Receptors (TLRs)

The TLR family of proteins is highly conserved during evolution and they have the capacity to recognize and bind with both self and non-self-molecules, which elicit both innate and adaptive responses. TLRs are unique in their ligand binding specificity and display distinct sensing and intracellular pathways. Among the TLR family of proteins, TLR4 can bind with a variety of Pathogen Associated Molecular Patterns (PAMPs) and Damage Associated Molecular patterns (DAMPs). The PAMPs of TLR4 includes gram-negative Lipopolysaccharide (LPS),<sup>25</sup> fibronectin,<sup>26</sup> hyaluronan,<sup>27</sup> fibrinogen,<sup>28</sup> heat shock proteins<sup>29</sup> and respiratory syncytial virus (RSV) fusion (F) protein, etc.<sup>30</sup> In humans and mice 13 toll-like receptors (TLRs) have been discovered. Nine TLRs are common between humans and mice; TLR10 is not present in mice and is expressed in humans. TLR11, 12 and 13 are expressed in mice and not expressed in human.<sup>30</sup> They are also termed as evolutionarily conserved receptors of the immune system.<sup>3,31</sup>

TLR stimulation is tightly controlled by phosphorylation events, ubiquitylation, physical interactions, conformational changes and proteasome-mediated degradation that are under the control of various regulatory molecules.<sup>3,31</sup> TLR4 gene is located in human chromosome 9 (9q32-q33) with 4 exons and 3 introns and of 19kb in sequence.<sup>15,32</sup> It is observed that single nucleotide polymorphisms in TLR4 gene can lead to the hypo-responsiveness for mounting immune reactions and pose the risk for various diseases which are inflammatory in nature including, atherosclerosis, Crohn's disease, ulcerative colitis, rheumatoid arthritis and prostate cancer.<sup>33,34,35</sup>

### TLR4 and Neuroinflammation

Neuroinflammation is the phenomenon of coordinated insult to the cells of the nervous system leading to tissue damage. Expression and activation of TLR4 lead to various inflammatory diseases. TLR4 can be activated by both exogenous and endogenous ligands and once activated it leads to robust production of proinflammatory cytokines.<sup>34,35</sup> TLRs are not only involved in the PAMPs and DAMPs induced neuroinflammatory changes in the brain, but they actively participate in the development of Parkinson's disease

(PD), amyotrophic lateral sclerosis (ALS) and Alzheimer's disease (AD).<sup>36</sup>

Various studies have reported the involvement of TLR4 in brain diseases. The signaling pathway studies showed that treatment of LPS in rat hippocampal neurons caused neuroinflammation through PTEN/PI3K/AKT/NF- $\kappa$ B pathway.<sup>37</sup> This study confirms the involvement of NF- $\kappa$ B in TLR4-LPS axis-induced neuroinflammation in the brain.

Inhibitory studies shed light on the role of TLR4 in neuroinflammation. Blockade of TLR4 signal inhibitor resatorvid during ischemic stress downregulated the phospho-p38, NF- $\kappa$ B, matrix metalloproteinase9 (MMP9) levels and also it reduced NOX4 expression thus preventing both nitrate and oxidative stress. This inhibition protected the neurons from apoptosis.<sup>38</sup> Curcumin inhibits the TLR4-MyD88-NF- $\kappa$ B axis and protects the brain from acute inflammatory injury during experimental traumatic brain injury.<sup>13</sup>

Some reports suggest that external antitumor agents used to treat cancers can exert their toxic effect on neuronal cells, through TLRs. Recently, it has been shown that, Paclitaxel, a chemotherapeutic agent, when administered at high concentrations through TLR4 mediates its damaging effects on axonal growth on DRG (dorsal root ganglion) neurons. The TLRs are expressed in CNS both in Glia and neurons and can bind with both exogenous as well as endogenous ligands.<sup>39</sup>

Specifically, TLRs are targeted as therapeutic agents in CNS inflammation.<sup>39</sup> In the case of CNS, TLR2 and TLR4 are reported to bind with DAMPs and PAMPs.<sup>39</sup> Various brain research studies reported the involvement of TLR4 as an important mediator of neuroinflammation during neuropathic pain<sup>40</sup> and in acute and chronic itch conditions. Moreover, alcohol-induced brain damage and neuroinflammation are mediated by TLR4.<sup>41,42</sup> The role of TLR4 in various neuronal injuries remains completely unknown and less understood.

### Cigarette Smoke and Neurological Disturbances

Cigarette smoke increases anxiety disorders and symptoms<sup>22</sup> and causes disturbances in blood-brain barrier BBB.<sup>44</sup> Oxidative stress and inflammation in rat brain have been the hallmark of cigarette smoke exposure.<sup>8,28</sup> Cigarette smoke also causes lipid peroxidation and increased the activity of acetylcholinesterase and these were ameliorated by vitamin E supplements.<sup>45</sup> Carbon monoxide in cigarette smoke disturbs the long-term potentiation in the hippocampus in rats.<sup>45</sup> Human smokers have both disturbed and poor quality of sleep followed by visual memory impairments when compared with non-smokers.<sup>4</sup> Structural abnormality in the brain and also neuropsychological tests have been noted in patients who are smokers as well as alcoholics.<sup>23</sup>

Nicotine had a profound impact on the attention index in human patients.<sup>27</sup> Grey matter volume and grey matter density was reduced in smokers when compared with non-smokers.<sup>46,47</sup> Such differences are noted in areas of the brain which mediate working memory, attention and drug reinforcement. Pregnant rats infused with Nicotine showed teratogenic effects and synaptic dysfunctions.<sup>48</sup>

Previous studies have shown that nicotine can cause increased release of glutamate, GABA, norepinephrine, dopamine and acetylcholine.<sup>49,50,51</sup> But the precise mechanism of such release is not yet understood. A study by Karama et al., 2015,<sup>29</sup> showed that in the human brain cortical thinning occurs because of Cigarette smoke. A total of 504 subjects were analyzed by brain magnetic resonance imaging (MRI) revealed this cortex thinning. Way back in 1998 the effects of cigarette smoke on working memory started to be noticed.<sup>29</sup> There are reports which suggest a protective role of nicotine in the brain, but the majority of studies reveal the negative effects of cigarette smoke on brain functions. Cigarette smoke has been associated with numerous health effects but in particular to the brain, as shown by the Centers for Disease Control and Prevention. Cognitive flexibility and memory are very poor in smokers.<sup>52</sup> There are reports which suggest that dementia and cigarette smoking to the extent of 14 % are associated with the development of Alzheimer's.<sup>53,54</sup>

Selected regions of the cortex of the smoking brain showed atrophy and reduced cortical volume.<sup>55-60</sup> Further, interesting studies report that in the frontal lobe there is a reduction in thickness and also abnormal levels of NAA, a marker for neuronal integrity (N-acetyl aspartate) was abnormal. This area of the brain is involved in addictive brain disorders.<sup>57,59</sup> Cigarette smoke causes cognitive impairment and also redox imbalance.<sup>61</sup> Medial orbito-frontal cortex showed a reduced thickness in smokers. This region is involved in the brain's reward, impulse control and decision-making. Cortical thinning is important because it is one of the markers for cognitive aging. Mojtaba et al., 2012<sup>62</sup> noted that cortical thinning is noted even after the cessation of Cigarette Smoke even after many years. The effects of cigarette smoke include cancer, neurological disorders, silent cerebral infarction, stroke and cerebral aneurysm.<sup>63-65</sup> Cigarettes which contain very low levels of nicotine as well as nicotine-free cigarettes are equally harmful to the blood-brain barrier endothelial cells.<sup>44</sup> Various disorders such as Alzheimer's, multiple sclerosis, small vessel ischemic disease, developmental problems during the pregnancy, all the other problems associated with cigarette smoking.<sup>66</sup> One more possibility that the nicotine present in cigarette smoke can activate the nicotinic receptors. Nicotine decreases the ZO1 expression, a tight junctional protein and leading to disturbances in ion homeostasis. In these

disturbances, the released product might activate the glial and neuronal cells for further exacerbation of the situation. The exact mechanism of action of cigarette smoke in brain neuroimmune/neuropathology is not yet understood. Recent studies point out the link between cigarette smoke and neurological disturbances.<sup>19,67</sup> Animal studies also reveal that exposure to cigarette smoke in general, can increase ROS levels and circulating pro-inflammatory markers and decreased levels of antioxidant markers.<sup>6,18,28,68-70</sup> In human smokers, urine levels of oxidation products and also immunosuppression are noted.<sup>18,68,69</sup>

In Lewis rat brain (Khanna et al., 2015)<sup>8</sup> found increased markers of proinflammatory cytokines and oxidative stress markers in the brain. This study explored the entire brain homogenates, but did not examine individual brain regions of cigarette smoke-exposed rats. The pro-inflammatory cytokines such as Tumor Necrosis Factor (TNF- $\alpha$ ), Interleukins-IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, IL-23, IL-17, IL-10 and Transforming Growth Factor (TGF- $\beta$ ) observed in this study can only be produced by the activation of TLR4 signaling pathways. This is also not examined yet and probably this is the first study that addresses this issue. The LPS in cigarette smoke may have activated TLR4 in the brain or the endogenous TLR4 ligands such as HMGB1, HSP60, 70 and 90 could have activated the TLR4 in microglial cells and the activated products from activated microglial cells could have further stimulated the inflammatory changes in brain regions to further exacerbate the already existing situation. The role of TLR4 in cigarette smoke-induced memory dysfunctions is not yet understood. The brain is a highly vulnerable organ to oxidative stress as it contains a high concentration of Polyunsaturated fatty acids (PUFAs). The free radicals present in cigarette smoke can easily damage the BBB and damage the PUFAs of neuronal cells. Cigarette Smoke contains biologically active endotoxin LPS, it can also be absorbed from the lungs and it can be seen in venous blood and in plasma by increasing the expression of TNF-, IL-6, sTNFR-I and II.<sup>2,45,71</sup>

In mice treated with alcohol, it showed disturbances in behavioral and cognitive disturbances whereas TLR-/- mice were protected from these effects during alcohol exposure.<sup>42</sup> LPS treated TLR-/- mice/rat is protected from brain inflammatory changes and cognitive dysfunctions. LPS is a component of tobacco smoke, dust and pollution, it is represented in various inflammatory diseases.<sup>2,45,71,72</sup> LPS is known to activate the brain neuronal and glial TLR4.<sup>73,74</sup> The pro-inflammatory cytokines such as Tumor Necrosis Factor (TNF- $\alpha$ ), Interleukins-IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, IL-23, IL-17, IL-10 and Transforming Growth Factor (TGF- $\beta$ ) observed in this study can only be produced by the activation of TLR4 signaling pathways. This is also not examined yet and probably this is the first study that addresses this issue.

## Discussion

Cigarette Smoke is injurious to health because it contains over 4,700 different chemical compounds which include oxidants and carcinogenic substances.<sup>11,16,38</sup> Cigarette smoke is formed of many toxic chemicals such as carcinogenic compounds<sup>41,42</sup> and also the free radicals<sup>18</sup> which can cause potent oxidative damage to tissues.<sup>19</sup> According to a scientific report death by tobacco increased from 5.4 million in 2005 and in 2006 it increased to 6.4 million and in 2030 it will increase to more than 8.3 million.<sup>20</sup> Death by tobacco accounts for 10% globally and it can kill almost 50% more people than the dangerous disease like HIV –AIDS.<sup>20</sup>

Evolution did not allow the cells of the human body to tolerate such toxic chemical compounds to pass on the cells without causing any notifiable injury to the cells. Lungs are very special organs because they do interact with the external environment through the epithelial cells. In particular, tobacco smoke, Cigarette Smoke and airborne pollutants can easily attack the lung epithelial cells, leading to inflammation. The consequences of such stress are the formation of oxidative stress, apoptosis, senescence and inflammation, increased HDAC activity, aggregation of insoluble polyubiquitinated aggregates and lack of antioxidant production.<sup>43,22</sup> Even though the lungs are special organs for counteracting pollutants and other toxic substances, Central Nervous System (CNS), especially the brain is highly vulnerable to these effects for the following reasons.

## Conclusion

The recent findings of TLR in Neurons, showed the involvement of external stimuli such as Cigarette smoke causing various neurological disorders and brain inflammation. The role of TLR4 in activating the brain during acute and chronic cigarette exposure is not yet investigated and also the neuroimmune mechanisms of cigarette smoke are not yet clearly understood. Smoking-related neurological disorders require proper understanding of the mechanisms of such damage for therapeutic development. Hence research should be carried out to study the TLR4 mechanisms and pathways in various inflammatory diseases.

## Acknowledgment

The authors are grateful for all the efforts made for this review.

**Conflict of Interest:** None

**Source of Funding:** None

## References

- Shukitt-Hale B, Stillman MJ, Lieberman HR. Tyrosine administration prevents hypoxia-induced decrements in learning and memory. *Physiol Behav.* 1996 Apr-May;59(4-5):867-71.[PubMed] [Google Scholar]
- Goto H, Rylander R. Kinetics of inhaled lipopolysaccharide in the guinea pig. *J Lab Clin Med.* 1987 Sep;110(3):287-91. [PubMed] [Google Scholar]
- Takeda K, Kaisho T and Akira S. Toll-like receptors. *Annu Rev Immunol.* 2003;21:335-76.
- Liu JT, Lee IH, Wang CH, Chen KC, Lee CI, Yang YK. Cigarette smoking might impair memory and sleep quality. *J Formos Med Assoc.* 2013 May;112(5):287-90. [PubMed] [Google Scholar]
- Gorina R, Font-Nieves M, Márquez-Kisinousky L, Santalucia T, Planas AM. Astrocyte TLR4 activation induces a proinflammatory environment through the interplay between MyD88-dependent NFκB signaling, MAPK and Jak1/Stat1 pathways. *Glia.* 2011 Feb;59(2):242-55. [PubMed] [Google Scholar]
- Moylan S, Jacka FN, Pasco JA, Berk M. How cigarette smoking may increase the risk of anxiety symptoms and anxiety disorders: A critical review of biological pathways. *Brain Behav.* 2013 May;3(3):302-26. [PubMed] [Google Scholar]
- Martínez-Cerdeño V, Noctor SC. Neural progenitor cell terminology. *Front Neuroanat.* 2018 Dec;12:104. [PubMed] [Google Scholar]
- Khanna A, Guo M, Mehra M, Royal W 3rd. Inflammation and oxidative stress induced by cigarette smoke in Lewis rat brains. *J Neuroimmunol.* 2013 Jan;254(1-2):69-75.[PubMed] [Google Scholar]
- Molteni M, Gemma S, Rossetti C. The role of toll-like receptor 4 in infectious and noninfectious inflammation. *Mediators Inflamm.* 2016;2016:6978936.[PubMed] [Google Scholar]
- Krishnan J, Selvarajoo K, Tsuchiya M, Lee G, Choi S. Toll-like receptor signal transduction. *Exp Mol Med.* 2007 Aug;39(4):421-38.[PubMed] [Google Scholar]
- Watson MB, Costello DA, Carney DG, McQuillan K, Lynch MA. SIGIRR modulates the inflammatory response in the brain. *Brain Behav Immun.* 2010 Aug;24(6):985-95. [PubMed] [Google Scholar]
- Watson SA, McStay GP. Functions of Cytochrome c oxidase assembly factors. *Int J Mol Sci.* 2020 Sep;21(19):7254.[PubMed] [Google Scholar]
- Zhu HT, Bian C, Yuan JC, Chu WH, Xiang X, Chen F, Wang CS, Feng H, Lin JK. Curcumin attenuates acute inflammatory injury by inhibiting the TLR4/MyD88/NF-κB signaling pathway in experimental traumatic brain injury. *J Neuroinflammation.* 2014 Mar;11:59. [PubMed] [Google Scholar]
- Manickam M, Tulsawani R. Survival response of hippocampal neurons under low oxygen conditions induced by Hippophae rhamnoides is associated with JAK/STAT signaling. *PLoS One.* 2014 Feb;9(2):e87694.

- [PubMed] [Google Scholar]
15. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell*. 2006 Feb;124(4):783-801. [PubMed] [Google Scholar]
  16. Keith CR, Thomann RV. Aerosol studies of cigarette smoke. *Environ Sci Technol*.1989;23:413-7.
  17. Genbacev-Krtolica O. Highlight for phenols, quinolines, indoles, benzene and 2-cyclopenten-1-ones are oviduct toxicants in cigarette smoke, by Prue Talbot, Karen Riveles and Ryan Rosa: List of tobacco-smoke constituents that are harmful for reproduction grows - Passive smoke. *Toxicol Sci*. 2005 Jul;86(1):4-5.[PubMed] [Google Scholar]
  18. Pryor WA, Prier DG, Church DF. Electron-spin resonance study of mainstream and sidestream cigarette smoke: Nature of the free radicals in gas-phase smoke and in cigarette tar. *Environ Health Perspect*. 1983 Jan;47:345-55. [PubMed] [Google Scholar]
  19. Moriarty SE, Shah JH, Lynn M, Jiang S, Openo K, Jones DP, Sternberg P. Oxidation of glutathione and cysteine in human plasma associated with smoking. *Free Radic Biol Med*. 2003 Dec;35(12):1582-8. [PubMed] [Google Scholar]
  20. Mathers CD, Loncar D.Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Med*. 2006 Nov;3(11):e442. [PubMed] [Google Scholar]
  21. Olsson B, Bondesson E, Borgström L, Edsbäcker S, Eirefelt S, Ekelund K, Gustavsson L, Hegelund-Myrbäck T. Pulmonary drug metabolism, clearance and absorption. In *Controlled pulmonary drug delivery 2011* (pp. 21-50). Springer, New York, NY. [Google Scholar]
  22. Van der Toorn M, Smit-de Vries MP, Slebos DJ, de Bruin HG, Abello N, van Oosterhout AJ, Bischoff R, Kauffman HF. Cigarette smoke irreversibly modifies glutathione in airway epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2007 Nov;293(5):L1156-62.[PubMed] [Google Scholar]
  23. Luppi, F., Aarbiou, J., van Wetering, S., Rahman, I., de Boer, W. I., Rabe, K. F., & Hiemstra, P. S. (2005). Effects of cigarette smoke condensate on proliferation and wound closure of bronchial epithelial cells in vitro: Role of glutathione. *Respiratory Research*, 6. [PubMed] [Google Scholar]
  24. Ransohoff RM, Brown MA. Innate immunity in the central nervous system. *J Clin Invest*. 2012 Apr;122(4):1164-71.[PubMed] [Google Scholar]
  25. Kawai T, Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity*. 2011 May;34(5):637-50. [PubMed] [Google Scholar]
  26. Okamura Y, Watari M, Jerud ES, Young DW, Ishizaka ST, Rose J, Chow JC, Strauss JF 3rd.The extra domain A of fibronectin activates toll-like receptor 4. *J Biol Chem*. 2001 Mar;276(13):10229-33.[PubMed] [Google Scholar]
  27. Jiang D, Liang J, Fan J, Yu S, Chen S, Luo Y, Prestwich GD, Mascarenhas MM, Garg HG, Quinn DA, Homer RJ, Goldstein DR, Bucala R, Lee PJ, Medzhitov R, Noble PW. Regulation of lung injury and repair by Toll-like receptors and hyaluronan. *Nat Med*. 2005 Nov;11(11):1173-9. [PubMed] [Google Scholar]
  28. Khanna AK, Xu J, Uber PA, Burke AP, Baquet C, Mehra MR.Tobacco smoke exposure in either the donor or recipient before transplantation accelerates cardiac allograft rejection, vascular inflammation and graft loss. *Circulation*. 2009 Nov;120(18):1814-21. [PubMed] [Google Scholar]
  29. Bulut Y, Faure E, Thomas L, Karahashi H, Michelsen KS, Equils O, Morrison SG, Morrison RP, Arditi M. Chlamydial heat shock protein 60 activates macrophages and endothelial cells through toll-like receptor 4 and MD2 in a MyD88-Dependent Pathway. *J Immunol*. 2002 Feb;168(3):1435-40.[PubMed] [Google Scholar]
  30. Kurt-Jones EA, Popova L, Kwinn L, Haynes LM, Jones LP, Tripp RA, Walsh EE, Freeman MW, Golenbock DT anderson LJ, Finberg RW. Pattern recognition receptors TLR4 and CD14 mediate response to respiratory syncytial virus. *Nat Immunol*. 2000 Nov;1(5):398-401.. [PubMed] [Google Scholar]
  31. Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol*. 2001 Aug;2(8):675-80.[PubMed] [Google Scholar]
  32. Horie Y, Meguro A, Ota M, Kitaichi N, Katsuyama Y, Takemoto Y, Namba K, Yoshida K, Song YW, Park KS, Lee EB, Inoko H, Mizuki N, Ohno S. Association of TLR4 polymorphisms with Behçet's disease in a Korean population. *Rheumatology (Oxford)*. 2009 Jun;48(6):638-42. [PubMed] [Google Scholar]
  33. Arbour NC, Lorenz E, Schutte BC, Zabner J, Kline JN, Jones M, Frees K, Watt JL, Schwartz DA. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet*. 2000 Jun;25(2):187-91. [PubMed] [Google Scholar]
  34. Schoch CL, Seifert KA, Huhndorf S, Robert V, Spouge JL, Levesque CA, Chen W; Fungal Barcoding Consortium; Fungal Barcoding Consortium Author List. Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proc Natl Acad Sci U S A*. 2012 Apr;109(16):6241-6.[PubMed] [Google Scholar]
  35. Schmitt C, Humeny A, Becker CM, Brune K, Pahl A. Polymorphisms of TLR4: Rapid genotyping and reduced response to lipopolysaccharide of TLR4 mutant alleles.

- Clin Chem. 2002 Oct;48(10):1661-7. [PubMed] [Google Scholar]
36. Heneka MT, Sastre M, Dumitrescu-Ozimek L, Dewachter I, Walter J, Klockgether T, Van Leuven F. Focal glial activation coincides with increased BACE1 activation and precedes amyloid plaque deposition in APP[V717I] transgenic mice. *J Neuroinflammation*. 2005 Oct;2:22. [PubMed] [Google Scholar]
  37. Zhao M, Zhou A, Xu L, Zhang X. The role of TLR4-mediated PTEN/PI3K/AKT/NF- $\kappa$ B signaling pathway in neuroinflammation in hippocampal neurons. *Neuroscience*. 2014 Jun;269:93-101. [PubMed] [Google Scholar]
  38. Suzuki Y, Hattori K, Hamanaka J, Murase T, Egashira Y, Mishiro K, Ishiguro M, Tsuruma K, Hirose Y, Tanaka H, Yoshimura S, Shimazawa M, Inagaki N, Nagasawa H, Iwama T, Hara H. Pharmacological inhibition of TLR4-NOX4 signal protects against neuronal death in transient focal ischemia. *Sci Rep*. 2012;2:896. [PubMed] [Google Scholar]
  39. Vega-Avelaira D, Géranton SM, Fitzgerald M. Differential regulation of immune responses and macrophage/neuron interactions in the dorsal root ganglion in young and adult rats following nerve injury. *Mol Pain*. 2009 Dec;5:70. [PubMed] [Google Scholar]
  40. Qi J, Buzas K, Fan H, Cohen JJ, Wang K, Mont E, Klinman D, Oppenheim JJ, Howard OM. Painful pathways induced by TLR stimulation of dorsal root ganglion neurons. *J Immunol*. 2011 Jun;186(11):6417-26. [PubMed] [Google Scholar]
  41. Alfonso-Loeches S, Pascual-Lucas M, Blanco AM, Sanchez-Vera I, Guerri C. Pivotal role of TLR4 receptors in alcohol-induced neuroinflammation and brain damage. *J Neurosci*. 2010 Jun;30(24):8285-95. [PubMed] [Google Scholar]
  42. Pascual M, Baliño P, Alfonso-Loeches S, Aragón CM, Guerri C. Impact of TLR4 on behavioral and cognitive dysfunctions associated with alcohol-induced neuroinflammatory damage. *Brain Behav Immun*. 2011 Jun;25 Suppl 1:S80-91. [PubMed] [Google Scholar]
  43. Hedström AK, Hillert J, Olsson T, Alfredsson L. Smoking and multiple sclerosis susceptibility. *Eur J Epidemiol*. 2013 Nov;28(11):867-74. [PubMed] [Google Scholar]
  44. Naik P, Fofaria N, Prasad S, Sajja RK, Weksler B, Couraud PO, Romero IA, Cucullo L. Oxidative and pro-inflammatory impact of regular and denicotinized cigarettes on blood brain barrier endothelial cells: is smoking reduced or nicotine-free products really safe? *BMC Neurosci*. 2014 Apr;15:51. [PubMed] [Google Scholar]
  45. Hasday JD, Bascom R, Costa JJ, Fitzgerald T, Dubin W. Bacterial endotoxin is an active component of cigarette smoke. *Chest*. 1999 Mar;115(3):829-35. [PubMed] [Google Scholar]
  46. Kühn S, Schubert F, Gallinat J. Reduced thickness of medial orbitofrontal cortex in smokers. *Biol Psychiatry*. 2010 Dec;68(11):1061-5. [PubMed] [Google Scholar]
  47. Gallinat J, Meisenzahl E, Jacobsen LK, Kalus P, Bierbrauer J, Kienast T, Witthaus H, Leopold K, Seifert F, Schubert F, Staedtgen M. Smoking and structural brain deficits: a volumetric MR investigation. *Eur J Neurosci*. 2006 Sep;24(6):1744-50. [PubMed] [Google Scholar]
  48. Vaglenova J, Parameshwaran K, Suppiramaniam V, Breese CR, Pandiella N, Birru S. Long-lasting teratogenic effects of nicotine on cognition: gender specificity and role of AMPA receptor function. *Neurobiol Learn Mem*. 2008 Oct;90(3):527-36. [PubMed] [Google Scholar]
  49. Ochoa EL, Li L, McNamee MG. Desensitization of central cholinergic mechanisms and neuroadaptation to nicotine. *Mol Neurobiol*. 1990 Fall-Winter;4(3-4):251-87. [PubMed] [Google Scholar]
  50. Léna C, Changeux JP, Mulle C. Evidence for "preterminal" nicotinic receptors on GABAergic axons in the rat interpeduncular nucleus. *J Neurosci*. 1993 Jun;13(6):2680-8. [PubMed] [Google Scholar]
  51. Henningfield JE, Cohen CA, Pickworth WB. Psychopharmacology of nicotine. *Nicotine addiction: Principles and management*. 1993:24-45. [Google Scholar]
  52. Corley J, Gow AJ, Starr JM, Deary IJ. Smoking, childhood IQ and cognitive function in old age. *J Psychosom Res*. 2012 Aug;73(2):132-8. [PubMed] [Google Scholar]
  53. Barnes PJ. New molecular targets for the treatment of neutrophilic diseases. *J Allergy Clin Immunol*. 2007 May;119(5):1055-62; quiz 1063-4. [PubMed] [Google Scholar]
  54. Barnes DE, Yaffe K. The projected effect of risk factor reduction on Alzheimer's disease prevalence. *Lancet Neurol*. 2011 Sep;10(9):819-28. [PubMed] [Google Scholar]
  55. Brody AL, Mandelkern MA, Jarvik ME, Lee GS, Smith EC, Huang JC, Bota RG, Bartzokis G, London ED. Differences between smokers and nonsmokers in regional gray matter volumes and densities. *Biol Psychiatry*. 2004 Jan;55(1):77-84. [PubMed] [Google Scholar]
  56. Almeida OP, Garrido GJ, Lautenschlager NT, Hulse GK, Jamrozik K, Flicker L. Smoking is associated with reduced cortical regional gray matter density in brain regions associated with incipient Alzheimer disease. *Am J Geriatr Psychiatry*. 2008 Jan;16(1):92-8. [PubMed] [Google Scholar]
  57. Durazzo TC, Mon A, Gazdzinski S, Meyerhoff DJ. Chronic cigarette smoking in alcohol dependence: associations with cortical thickness and N-acetylaspartate levels in

- the extended brain reward system. *Addict Biol.* 2013 Mar;18(2):379-91. [PubMed] [Google Scholar]
58. Durazzo TC, Cardenas VA, Studholme C, Weiner MW, Meyerhoff DJ. Non-treatment-seeking heavy drinkers: effects of chronic cigarette smoking on brain structure. *Drug Alcohol Depend.* 2007 Feb;87(1):76-82. [PubMed] [Google Scholar]
59. Durazzo TC, Insel PS, Weiner MW; Alzheimer Disease Neuroimaging Initiative. Greater regional brain atrophy rate in healthy elderly subjects with a history of cigarette smoking. *Alzheimers Dement.* 2012 Nov;8(6):513-9. [PubMed] [Google Scholar]
60. Akiyama H, Meyer JS, Mortel KF, Terayama Y, Thornby JI, Konno S. Normal human aging: factors contributing to cerebral atrophy. *J Neurol Sci.* 1997 Nov;152(1):39-49. [PubMed] [Google Scholar]
61. Pierik M, Vermeire S, El-Housni H, Claessens G, Quertinmont E, Joosens S, Van Gossun A, Deviere J, Rutgeerts P, Franchimont D. Toll-like receptor (TLR)-4 Asp299gly polymorphism is associated with ulcerative colitis (UC) O. *Gastroenterology.* 2003;4(124):A370. [Google Scholar]
62. Zarei M, Ibarretxe-Bilbao N, Compta Y, Hough M, Junque C, Bargallo N, Tolosa E, Martí MJ. Cortical thinning is associated with disease stages and dementia in Parkinson's disease. *J Neurol Neurosurg Psychiatry.* 2013 Aug;84(8):875-81. [PubMed] [Google Scholar]
63. Miller GJ, Bauer KA, Cooper JA, Rosenberg RD. Activation of the coagulant pathway in cigarette smokers. *Thromb Haemost.* 1998 Mar;79(3):549-53. [PubMed] [Google Scholar]
64. Satoto TBT, Satrisno H, Lazuardi L, Diptyanusa A, Purwaningsih, Rumbiwati, Kuswati. Insecticide resistance in *Aedes aegypti*: An impact from human urbanization? *PLoS One.* 2019 Jun 24;14(6):e0218079. [PubMed] [Google Scholar]
65. Rahman SM, Hossain SM. Dengue prevention and control: Bangladesh context: Dengue Bangladesh Context. *Bangladesh Med Res Counc Bull.* 2019 Aug;45(2):66-7. [Google Scholar]
66. Chan A, Chiang LP, Hapuarachchi HC, Tan CH, Pang SC, Lee R, Lee KS, Ng LC, Lam-Phua SG. DNA barcoding: complementing morphological identification of mosquito species in Singapore. *Parasit Vectors.* 2014 Dec;7:569. [PubMed] [Google Scholar]
67. Shah RS, Cole JW. Smoking and stroke: the more you smoke the more you stroke. *Expert Rev Cardiovasc Ther.* 2010 Jul;8(7):917-32. [PubMed] [Google Scholar]
68. Harman SM, Liang L, Tsitouras PD, Gucciardo F, Heward CB, Reaven PD, Ping W, Ahmed A, Cutler RG. Urinary excretion of three nucleic acid oxidation adducts and isoprostane F(2)alpha measured by liquid chromatography-mass spectrometry in smokers, ex-smokers and nonsmokers. *Free Radic Biol Med.* 2003 Nov;35(10):1301-9. [PubMed] [Google Scholar]
69. Chang RC, Ho YS, Wong S, Gentleman SM, Ng HK. Neuropathology of cigarette smoking. *Acta Neuropathol.* 2014 Jan;127(1):53-69. [PubMed] [Google Scholar]
70. Chen H, Cowan MJ, Hasday JD, Vogel SN, Medvedev AE. Tobacco smoking inhibits expression of proinflammatory cytokines and activation of IL-1R-associated kinase, p38 and NF-kappaB in alveolar macrophages stimulated with TLR2 and TLR4 agonists. *J Immunol.* 2007 Nov;179(9):6097-106. [PubMed] [Google Scholar]
71. Michie HR, Manogue KR, Spriggs DR, Revhaug A, O'Dwyer S, Dinarello CA, Cerami A, Wolff SM, Wilmore DW. Detection of circulating tumor necrosis factor after endotoxin administration. *N Engl J Med.* 1988 Jun;318(23):1481-6. [PubMed] [Google Scholar]
72. Larsson L, Szponar B, Pehrson C. Tobacco smoking increases dramatically air concentrations of endotoxin. *Indoor Air.* 2004 Dec;14(6):421-4. [PubMed] [Google Scholar]
73. Hines DJ, Choi HB, Hines RM, Phillips AG, MacVicar BA. Prevention of LPS-induced microglia activation, cytokine production and sickness behavior with TLR4 receptor interfering peptides. *PLoS One.* 2013;8(3):e60388. [PubMed] [Google Scholar]
74. Leow-Dyke S, Allen C, Denes A, Nilsson O, Maysami S, Bowie AG, Rothwell NJ, Pinteaux E. Neuronal Toll-like receptor 4 signaling induces brain endothelial activation and neutrophil transmigration in vitro. *J Neuroinflammation.* 2012 Oct;9:230. [PubMed] [Google Scholar]