

SPECIAL ARTICLE

A Brief History of the Purified Chick Embryo Cell Human Rabies Vaccine, ChiroRab (formerly known as Rabipur)

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ABSTRACT

The development of modern cell culture vaccines revolutionized human rabies prevention since the latter part of the 20th century. ChiroRab (previously known as Rabipur) is an inactivated purified chick embryo cell rabies vaccine, produced in India for the prevention of rabies in humans. Over the past 35 years, this pure, potent, safe and efficacious biologic, when administered by the intramuscular or intradermal routes, in pre- or postexposure prophylaxis, prevented rabies in millions of exposed people in Africa, the Americas, Australia, and Eurasia. Based upon its sound clinical performance to date, ChiroRab will continue to play a major role in the global elimination of human rabies caused from dogs by 2030, particularly throughout the Indian subcontinent.

INTRODUCTION

Rabies, an acute progressive encephalitis, is one of the oldest described infectious diseases – some believe its origins transcend human history (1-3). The etiological agents consist of single-stranded, negative-sense RNA viruses, with more than 17 recognized and putative viral species (4,5). Multiple lyssaviruses utilize bats and other mammals as reservoirs on all continents, except Antarctica (6,7). As quintessential neurotropic viruses, lyssaviruses exploit routine mammalian behaviors to ensure their transmission via the transdermal or mucosal routes (8-11). From the standpoint of concerns in agriculture, biological conservation and public health, rabies virus is the most important lyssavirus and dogs constitute the most prominent reservoir (8,12). Unlike diseases such as smallpox and rinderpest, rabies is not a candidate for eradication (13). However, within the 21st century, rabies can be prevented, controlled and in some cases, selectively eliminated from circulation, over large geographic regions (8,14-16).

Before the age of vaccination, rabies management included human avoidance of animal bites, leashing and muzzling of owned dogs, culling of stray dogs and a wide variety of techniques (e.g., amputation, cauterization, 'quackery', etc.) in an attempt to prevent a productive viral infection after a bite from a rabid animal (17). After Pasteur's historical demonstration of the first human rabies vaccination during the late 19th century, gradual improvements followed, towards further development of nerve tissue vaccines (NTV) produced in animals (e.g., rabbits, sheep, mice, etc.). Unfortunately, NTV suffered from issues related to lower potency and product safety (18). During the mid-20th century, use of avian embryos, primary cell cultures and continuous cell lines for defined viral propagation revolutionized rabies vaccine development from the era of the NTV predominance (Table 1).

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Table 1. Historical examples in the evolution of human rabies vaccines

Type	Substrate	Comments	References
Pasteur, 1885	Adult nervous tissue vaccine (NTV), produced in dried spinal cord of rabbits	Residual rabies virus, high myelin content, required > 14 doses administered subcutaneously in the abdominal area	17
Fermi	NTV	Partial chemical treatment of rabies virus-infected tissue specimens for vaccine preparation, with a combination of both inactivated and replication-competent virions	19
Semple	NTV	Greater inactivation of rabies virus-infected rabbit, sheep or goat tissues, initially by carbolic acid (i.e., phenol), with widespread use, particularly in India, but with residual risk of post-vaccinal encephalomyelitis	20
Hempt	NTV	Ether treatment prior to phenol inactivation and stabilization for distribution over broader geographic areas, especially throughout Central Europe, but with risk of post-vaccinal encephalomyelitis	21
Fuenzalida Palacios	Suckling mouse NTV	Safer than adult NTV, inactivated by ultra-violet (UV) irradiation (after infection with CVS rabies virus), once widely used throughout Latin America	22
Duck embryo vaccine (DEV)	Duck embryos	Infected with PM rabies virus strains, and inactivated by beta-propiolactone (BPL), hence safer than NTV, but poorly immunogenic	23
Primary hamster kidney cell vaccine (PHKC)	Young Syrian hamster kidney cell culture	UV-inactivation of cells infected with the Vnukovo rabies virus strain, used widely in China and the former USSR	24
Rabies vaccine adsorbed (RVA)	Diploid fetal rhesus monkey lung fibroblasts	BPL-treated rabies virus, adsorbed onto aluminum Phosphate, produced originally by the Michigan Department of Health, USA	25
Human diploid cell vaccine (HDCV)	WI-38 cells, later MRC-5 cells	Inactivated by BPL, much safer than NTV, highly potent, immunogenic and efficacious, used in multiple countries, especially in Australia, Europe and North America	26-28

Type	Substrate	Comments	References
Purified vero cell rabies vaccine (PVRV)	Vero cells	Rabies virus (PM strain) inactivated by BPL, much safer than NTV, highly potent, immunogenic, efficacious, and economical to produce for global use	29,30
Purified chick embryo cell vaccine (PCEC)	Chicken fibroblasts	Rabies virus (LEP strain) inactivated by BPL, much safer than NTV, highly potent, immunogenic, efficacious, and economical to produce for global use	31-33

Whereas the first duck embryo vaccine (DEV) was an important milestone in safety over the NTV progenitors, potency remained low (23,34). Later, at the Wistar Institute of Anatomy & Biology, the human diploid cell vaccine (HDCV) became the first modern cell culture vaccine with both high safety and potency (26-28). Despite its overall superiority to NTV and DEV, higher production costs of HDCV limited its availability largely to the developed world. This drawback was overcome in part by the effective production of a purified vero cell vaccine, PVRV (29,30). Similarly, by repurposing the utility of avian tissues for rabies virus propagation, a purified chick embryo cell vaccine (PCEC) joined the vanguard for economic human rabies prevention in the developing world (31-33, 35,36). These modern cell culture rabies vaccines resulted in far fewer serious adverse events than the NTV, were highly immunogenic in the rapid induction of rabies virus neutralizing antibodies (VNA), were suitable for delivery via the intramuscular (IM) or intradermal (ID) routes by a variety of regimens, and could be used for the preexposure (PrEP) immunization of persons at risk of occupational rabies virus exposure (e.g., laboratory workers, veterinarians, etc.) as well as for postexposure prophylaxis (PEP) for the greater population at large after rabies virus exposure (i.e., typically after a rabid animal bite). Since their advent, several different brands of HDCV, PVRV and PCEC vaccines were marketed (8). In the interest of clarity, the objective of this communique is to provide a brief history of the PCEC vaccine, Rabavert, known today as ChiroRab (Figure 1).

Figure 1 ChiroRab Rabies Vaccine (NB: please insert a better product image if available)



PURIFIED CHICK EMBRYO CELL VACCINE

The adaption of different rabies viruses, isolated from nature and serial passage within animals, primary cells or continuous cell cultures, serve as vaccine seed strains (e.g., CVS, PM, PV, ERA/SAD, etc.). The Flury rabies virus strain was isolated from a child infected through exposure to a rabid dog in Georgia during 1939 (37). This isolate was attenuated by serial brain passages within chicks and chick embryos (38). Historically, these were differentiated as the Flury low egg-passage virus

strain (i.e., LEP, < 50 passages) or the high-egg passage virus strain (i.e., HEP, > 179 passages). During the 1950-1960s, HEP and LEP rabies viruses were considered for use in both human and veterinary vaccine applications (39-44). Given the prior utility of the LEP virus propagated in avian tissues as a biologic, further investigations began during the late 1970s in Marburg, Germany, and during the 1980s, the PCEC vaccine was developed. In a series of pre-clinical tests, PCEC vaccine demonstrated comparable potency and safety to HDCV. Thereafter, in human clinical studies, PCEC vaccine showed similar immunogenicity to HDCV and was subsequently licensed in Germany during 1984.

Over the past 35 years, Rabipur (today known as ChiroRab), has been used effectively for both PrEP and PEP (Table 2). In a recent meta-analysis of PCEC vaccine, nearly all individuals had rabies VNA above the minimum WHO suggested value, as indicative of an adequate immune response to vaccination (i.e., ≥ 0.5 IU/ml by day 14) and in the vaccinated population at large, had an acceptable safety profile (36).

Table 2. Comparative clinical use of Rabipur (AKA today, ChiroRab)

Observation	Comments	References
Safety	Rabipur was well tolerated by human subjects in multiple studies in China, Europe, India, Thailand & the USA	35
PrEP	Kinetics, height of the response and persistence of VNA over two years were virtually identical after Rabipur and HDCV administration	45
Comparable immunogenicity	Rabipur was as effective as HDCV at inducing an antibody titre of > 0.5 IU/ml in all Indian subjects, irrespective of their age and sex	46
Duration of VNA after Zagreb regimen	All vaccinees had high VNA levels from day 14, which lasted more than a year and the adverse reactions of the Rabipur vaccine were mild and self-limiting	47
Concomitant ID vaccination and RIG administration	Healthy volunteers received either ID Rabipur vaccine alone (0.1 ml at each of two sites on days 0, 3 and 7, and at one site on days 28 and 90), or similar vaccine with one dose of HRIG IM on day 0 and VNA was detectable in all subjects, in both groups, from day 14 to day 365, but no significant suppressive effects of RIG were observed.	48
Vaccine volume	In this population of healthy volunteers, a full dose of Rabipur in a dilution of 0.5 mL was as well tolerated locally and systemically as in a dilution of 1.0 mL and all subjects developed levels of VNA exceeding 0.5 IU/mL	49
Safety and efficacy in Thai patients using a dose-sparing regimen	Rabipur was safe and highly immunogenic when administered ID in 0.1-ml doses using a two-site method (i.e., on days 2,2,2,0,1,1)	50
Efficacy after exposure to confirmed rabid animals	A multi-site regimen of Rabipur, with or without passive immunization, prevented the development of rabies encephalitis in patients bitten by confirmed rabid dogs, over a 3-year observation period	51
Comparative response of different regimens with or without immune globulin	All participants given Rabipur ID achieved adequate VNA concentrations regardless of regimen or co-administration of RIG, but VNA concentrations were consistently lower in adults with RIG administration than in those without	52
Comparability via regimen	Vaccination with Rabipur under a 2-1-1 regimen was as safe and immunogenic as under a traditional 5-dose Essen regimen in healthy adult Chinese and Indian subjects	53, 54
Longevity after ID vaccination	All subjects had positive VNA in excess of 1 year	55
PEP and ID administration	Rabipur was safe and effective during PEP by the ID route in 43 Indian dog bite cases	56

Observation	Comments	References
Abbreviated, ID regimen	All subjects, who had nursed or casually handled a rabies patient and administered Rabipur, experienced seroconversion and all developed VNA levels higher than the adequate level	57
Effect of reduced volume of vaccine by the ID route	Adequate antibody response obtained with Rabipur when administered by the TRC regimen, even after reducing the quantity of vaccine from 0.2 ml to 0.1 ml per intradermal dose	58
Immunogenicity by number of ID site administration	Both a 2-site and 8-site ID regimens with purified chick embryo cell vaccine produced adequate levels of VNA	59
Cost-effective, dose sparing efficacy	Rabipur efficacy confirmed using the TRC regimen in 113 exposed patients in the Philippines, where vaccine was well tolerated, no vaccine-related serious adverse events occurred, and all patients were alive at least 1 year after initial exposure	60
Comparison to the Thai Red Cross regimen in India	Rabipur was safe and highly immunogenic in Indian subjects when administered intradermally as 0.1 mL/site using the "2-2-2-0-1-1" PEP regimen	61
ID vaccination using the Kempegowda Institute of Medical Sciences (KIMS), Bangalore, India, regimen	In healthy volunteers, PEP with Rabipur, administered using the KIMS-ID regimen, was well tolerated and immunogenic for a year	62
Comparative utility to PVRV	Rabipur was immunogenic, efficacious, and well tolerated when administered in the TRC regimen in ID doses of 0.1 mL compared to PVRV	63
Relationship of potency to immune response	No significant linear relationship between antigenicity and immunogenicity of Rabipur compared to HDCV and PVRV vaccines, when administered by the intradermal route	64
Comparability with other CCV	Rabipur was found safe, tolerable, and immunogenic as PDEV and PVRV in exposed Indian patients	65
Effect of prior route of administration	Rabipur was safe and immunogenic following booster vaccination, after cross-over from the ID to the IM route and vice versa	66
Switches in vaccine brand during prophylaxis	PEP was safe and immunogenic, despite changes in the route of administration and brand/type of vaccine (e.g., Rabipur, PVRV, etc.)	67
Shortened regimens	In a clinical comparison of Rabipur and PVRV, a new "one week ID regimen" was both immunogenic and safe in healthy Indian volunteers	68
Comparison of vaccine booster number	Clinically insignificant benefit of using 1 vs. 2 boosters of PCEC vaccine	69
Anamnestic responses	Anamnestic responses to booster doses of Rabipur demonstrated at least 14 years after the last vaccination	70
Pregnancy	Rabipur well tolerated with no reports of systemic or local AEs and the exposed patients had normal deliveries of healthy babies with no evidence of congenital abnormalities	71
Effect of age and regimen	Vaccination with Rabipur following a Zagreb regimen induced immune responses non-inferior to those of the Essen regimen, and had a similar safety and tolerability profile to the Essen regimen in Chinese children, adolescents, and adults over 51 years	72

Observation	Comments	References
PrEP in children	Rabipur was safe and immunogenic when administered for PrEP in Thai and Indian children	73, 74
Concomitant administration with Japanese encephalitis vaccine	After concomitant administration of Rabipur and Japanese encephalitis vaccine to Thai toddlers, adequate rabies and Japanese encephalitis VNA were demonstrated by day 49, 7 days after a booster at 1 year, and in a majorly at 3 years postvaccination	75
Potential compatibility with other travel vaccines	Raipur and Japanese encephalitis vaccine, administered together, with or without a quadrivalent meningococcal glycoconjugate vaccine, did not compromise immunogenicity or safety of the individual products	76
Malnourished subjects	No significant difference in VNA between malnutrition categories identified (i.e., immune response to Rabipur was independent of the extent of malnutrition) and no SAEs reported, indicating that the vaccine was well tolerated	77
Immune-compromised patients	Rabipur was immunogenic in HIV-1-infected patients with CD4+ cell counts below 200 cells/microL when administered in a modified eight-site intradermal PEP regimen	78
Comparability by commercial site of vaccine production	Noninferiority in immunogenicity and tolerability of the Rabipur produced in India to German vaccine batches	79
Exposure to bat lyssaviruses	Sera from volunteers who received Rabipur in a simulated ID PEP regimen contained adequate neutralizing antibodies to European and Australian bat lyssaviruses	80

Based upon such extensive studies, one may summarize the clinical experience with this PCEC vaccine:

- ChiroRab, administered using the recommended WHO PrEP schedule (e.g., days 0, 7, 21 or 28), provides an adequate concentration of VNA at ≥ 0.5 IU/mL by day 28, or earlier, in all subjects.
- ChiroRab can be administered safely IM or ID, using various schedules for PrEP and PEP and as booster vaccinations, in all age groups.
- ChiroRab is highly immunogenic, as 98% to 100% of subjects achieved VNA concentrations ≥ 0.5 IU/mL after PEP on day 14, regardless of the administration route (IM or ID).
- ChiroRab is efficacious, with regards to 1-year survival (100% survival seen in >120 subjects).
- ChiroRab displays a rapid anamnestic response, even years after primary vaccination, providing increased immunity and VNA of ≥ 0.5 IU/mL.
- ChiroRab has an established safety profile, with a positive benefit-risk profile, and no documented cases of true vaccine failure (i.e., fatal rabies virus infection after correct PEP administration) were identified, in contrast to the few cases where infection after PEP occurred, because prophylaxis had been inadequate (i.e., not according to WHO recommendations).

BASIC DESCRIPTION OF CHIRORAB VACCINE

ChiroRab is a sterile, freeze-dried vaccine obtained by replicating the fixed-virus strain Flury LEP in primary cultures of chicken fibroblasts using specific pathogen-free eggs in compliance with current applicable pharmacopeia and with WHO requirements. The rabies virus seed strain Flury LEP was obtained from the American Type Culture Collection at the 59th egg passage. Propagation of the LEP rabies virus occurs in a synthetic cell culture growth medium, with the addition of human albumin, polygeline (processed bovine gelatin), and antibiotics. The LEP rabies virus is inactivated by β -propiolactone and purified via continuous density-gradient centrifugation. Lyophilization of the vaccine occurs after addition of a stabilizer

solution, consisting of buffered polygeline and potassium glutamate. A dose of the reconstituted vaccine contains ≤ 12 mg polygeline (processed bovine gelatin), ≤ 0.3 mg human serum albumin, 1 mg potassium glutamate, and 0.3 mg sodium EDTA. Small amounts of bovine serum are used in the cell culture process. The bovine components originate from Australia, New Zealand, and the USA. Minimal amounts of avian protein may be present in the final product. The ovalbumin content is ≤ 3 ng/dose (1 mL), based on ELISA testing. Antibiotics (e.g., neomycin, chlortetracycline, amphotericin B), added during cell and virus propagation, are largely removed during subsequent steps in the production process. Neomycin is present at ≤ 10 mcg, chlortetracycline at ≤ 200 ng, and amphotericin B at ≤ 20 ng per vaccine dose. The vaccine is intended for intramuscular (IM) or intradermal (ID) administration. The vaccine contains no preservative and should be used immediately after reconstitution with the supplied Sterile Diluent (Water for Injection). The potency of the final vaccine product is determined by the NIH test. The potency of 1 dose (1.0 mL) of vaccine is equal to at least 2.5 IU of rabies virus antigen. This biologic is a white, freeze-dried vaccine for reconstitution with a diluent prior to human parenteral use. The reconstituted vaccine is a clear to slightly opalescent, colorless to slightly pink suspension. Until recently, PCEC vaccine was produced at two WHO pre-qualified manufacturing facilities: Germany and India. Batches of vaccine from the two sites were clinically comparable (79). Today, ChiroRab is produced only in Ankleshwar, India. Besides being efficacious against rabies virus variants, VNA elicited after ChiroRab immunization also cross react against other lyssaviruses, which is important given the opportunity for their potential emergence in India (80,81)

Based in part upon global demand, product supply, regulatory compliance and SAGE recommendations, rabies as a vaccine preventable disease was listed as a high priority candidate by the WHO during 2018-2020, in which Rabipur (and in the near future, ChiroRab) played a prominent role

(<https://extranet.who.int/pqweb/vaccines/eligibility>).

ASCENDENCY OF CHIRORAB®

In the 1980s, Rabipur was manufactured in Marburg, Germany by Behringwerke (a constituent of the Hoechst Group) and was exclusively imported and marketed in India (marketed by Hoechst India Ltd., Bombay). During 1989, a facility was opened at Ankleshwar, Gujarat for manufacturing and production of Rabipur in India, in technical collaboration with Behringwerke. In 1996, the vaccine business of Behringwerke (including that of Rabipur® vaccine and its technology) was transferred to the Chiron Corporation, known as Chiron Behring GmbH & Company, Germany. Consequently, in India too, new entity M/s Chiron Behring Vaccines Private Limited (CBVPL) was incorporated to take over existing manufacturing facility (including technology) from Hoechst India Limited. Further during 1998, Novartis Group took over the Global Chiron Corporation, thereby CBPL became the subsidiary of Novartis Group. In 2014, Novartis sold its vaccine business to GlaxoSmithKline (GSK).

In 2019, Bharat Biotech acquired legal entity Chiron Behring Vaccines from GSK, along with all technology, rights regarding clinical development, manufacturing process, QC processes, distribution and sale of rabies vaccine. All licenses and regulatory approvals obtained for Rabipur® have been fully transferred to ChiroRab®, including WHO prequalification which is in process. Chiron Behring Vaccines is one of the largest manufacturers of rabies vaccines in the world.

As can be seen production of ARV has continued past 3 decades at same plant with same technology, although there has been change in managements / Trade name. Such emphasis on vaccine supply is critical, given international aspirations for rabies prevention and control. For example, during 2015, the WHO announced a program for the global elimination of human rabies caused by dogs (GEHRD) via mass canine vaccination and human rabies prophylaxis, according to an increased emphasis on PrEP together with new ID dose-sparing and shortened PEP regimens (82-89). Clinicians have a major responsibility in selection of the ideal rabies biologics for their patients, to prevent deaths from a zoonosis with the highest case fatality of any infectious disease and to remain alert for counterfeit vaccines (90). Considering its considerable public health benefits shown to date, ChiroRab's future legacy includes a prominent role in progressing toward the GEHRD by 2030, especially throughout the Indian subcontinent and the surrounding region.

References

1. Badrane H., Tordo N., Host switching in Lyssavirus history from the Chiroptera to the Carnivora orders. *J Virol.* 2001 Sep;75(17):8096-104.
2. Hayman D.T., Fooks A.R., Marston D.A., Garcia-R JC. The Global Phylogeography of Lyssaviruses - Challenging the 'Out of Africa' Hypothesis. *PLoS Negl Trop Dis.* 2016 Dec 30;10(12):e0005266.
3. Rupprecht C, Kuzmin I, Meslin F. Lyssaviruses and rabies: current conundrums, concerns, contradictions and controversies. *F1000Res.* 2017 Feb 23;6:184.

4. International Committee on Taxonomy of Viruses. Virus Taxonomy: 2019 Release. (https://talk.ictvonline.org/ictv-reports/ictv_online_report/negative-sense-rna-viruses/w/rhabdoviridae/795/genus-lyssavirus).
5. Grobler CS, Coertse J, Markotter W. Complete Genome Sequence of Matlo Bat Lyssavirus. *Microbiol Resour Announc*. 2021 May 20;10(20):e00241-21.
6. Troupin C, Dacheux L, Tanguy M, et al. Large-Scale Phylogenomic Analysis Reveals the Complex Evolutionary History of Rabies Virus in Multiple Carnivore Hosts. *PLoS Pathog*. 2016 Dec 15;12(12):e1006041.
7. Rupprecht CE, Bannazadeh Baghi H, Del Rio Vilas VJ, et al. Historical, current and expected future occurrence of rabies in enzootic regions. *Rev Sci Tech*. 2018 Aug;37(2):729-739.
8. World Health Organization, 2018. World Health Organization Expert Consultation on Rabies. Second Report. Technical Report Series 1012. Geneva, Switzerland.
9. Davis BM, Rall GF, Schnell MJ. Everything You Always Wanted to Know About Rabies Virus (But Were Afraid to Ask). *Annu Rev Virol*. 2015 Nov;2(1):451-71.
10. Jackson AC. Rabies: a medical perspective. *Rev Sci Tech*. 2018 Aug;37(2):569-580.
11. Rohde RE, Rupprecht CE. Update on lyssaviruses and rabies: will past progress play as prologue in the near term towards future elimination? *Fac Rev*. 2020 Nov 16;9:9.
12. Hampson K, Coudeville L, Lembo T, et al. Estimating the global burden of endemic canine rabies. *PLoS Negl Trop Dis*. 2015 Apr 16;9(4):e0003709.
13. Rupprecht CE, Barrett J, Briggs D, et al. Can rabies be eradicated? *Dev Biol (Basel)*. 2008;131:95-121.
14. Vigilato MAN, Molina-Flores B, Del Rio Vilas VJ, Pompei JC, Cosivi O. Canine rabies elimination: governance principles. *Rev Sci Tech*. 2018 Aug;37(2):703-709.
15. Robardet E, Bosnjak D, Englund L, Demetriou P, Martin PR, Cliquet F. Zero Endemic Cases of Wildlife Rabies (Classical Rabies Virus, RABV) in the European Union by 2020: An Achievable Goal. *Trop Med Infect Dis*. 2019 Sep 30;4(4):124.
16. Ma X, Monroe BP, Wallace RM, et al. Rabies surveillance in the United States during 2019. *J Am Vet Med Assoc*. 2021 Jun 1;258(11):1205-1220.
17. Tarantola A. Four Thousand Years of Concepts Relating to Rabies in Animals and Humans, Its Prevention and Its Cure. *Trop Med Infect Dis*. 2017 Mar 24;2(2):5.
18. Chakrabarti P. "Living versus dead": The Pasteurian paradigm and imperial vaccine research. *Bull Hist Med*. 2010 Fall;84(3):387-423.
19. Fermi C. Über die Immunisierung gegen Wutkrankheit. *Zeitschrift für Hygiene und Infektionskrankheiten*. 1908;58:233-276.
20. Semple D. The Preparation of a Safe and Efficient Antirabic Vaccine. *Scientific Memoirs by Officers of the Medical and Sanitary Departments of the Government of India; Calcutta, India: 1911, 1-32.*
21. Hempt A. Sur une methode rapide de traitement antirabique. *Ann Institut Pasteur Paris* 1925, 39(7):632-40.
22. Fuenzalida E, Palacios R, Borgono JM. Antirabies antibody response in man to vaccine made from infected suckling-mouse brains. *Bull World Health Organ* 1964;30:431-6.
23. Lavender JF, van Frank RM. Zonal-centrifuged purified duck embryo cell culture rabies vaccine for human vaccination. *Appl Microbiol* 1971;22:358-65.
24. Selimov M, Aksenova T, Klyueva E, Gribencha L, Lebedeva I. Evaluation of the inactivated tissue culture rabies vaccine from the Vnukovo-32 strain. Results of its industrial production and field use for post-exposure immunization of man. *Dev Biol Stand*. 1978;40:57-62.
25. Berlin BS, Mitchell JR, Burgoyne GH, et al. Rhesus diploid rabies vaccine (adsorbed), a new rabies vaccine. Results of initial clinical studies of preexposure vaccination. *JAMA*. 1982 Mar 26;247(12):1726-8.
26. Kuwert EK, Marcus I, Werner J, et al. Post-exposure use of human diploid cell culture rabies vaccine. *Dev Biol Stand*. 1976 Dec 13-15;37:273-86.

27. Wiktor TJ, Plotkin S, Koprowski H. Development and clinical trials of the new human rabies vaccine of tissue culture (human diploid cell) origin. *Dev Biol Stand.* 1978;40: 3–9.
28. Plotkin SA. Rabies vaccine prepared in human cell cultures: progress and perspectives. *Rev Infect Dis.* 1980 May-Jun;2(3):433-48.
29. Roumiantzeff M, Ajjan N, Branche, R, et al. Rabies vaccine produced in cell culture: production and control and clinical results. In: *Applied Virology* (ed. E. Kurstak), 1984, pp. 241-296. Orlando, Florida: Academic Press.
30. Toovey S. Preventing rabies with the Verorab vaccine: 1985-2005 Twenty years of clinical experience. *Travel Med Infect Dis* 2007;5:327-48.
31. Barth R, Bijok U, Grushkau H, Smerdel J, Vodopija J. Purified chicken embryo cell rabies vaccine for human use. *Lancet.* 1983 Mar 26;1(8326 Pt 1):700.
32. Barth R, Gruschkau H, Bijok U, et al. A new inactivated tissue culture rabies vaccine for use in man. Evaluation of PCEC-vaccine by laboratory tests. *J Biol Stand.* 1984 Jan;12(1):29-46.
33. Barth R, Gruschkau H, Jaeger O, Milcke L, Weinmann E. Purified chick embryo cell (PCEC) rabies vaccine for human use. Laboratory data. *Behring Inst Mitt.* 1984 Nov;(76):142-54.
34. Peck FG Jr, Powell HN, Culbertson CG. Duck embryo rabies vaccine. *JAMA.* 1956;162:1373-1376.
35. Giesen A, Gniel D, Malerczyk C. 30 Years of rabies vaccination with Rabipur: a summary of clinical data and global experience. *Expert Rev Vaccines.* 2015 Mar;14(3):351-67.
36. Preiss S, Chanthavanich P, Chen LH, et al. Post-exposure prophylaxis (PEP) for rabies with purified chick embryo cell vaccine: a systematic literature review and meta-analysis. *Expert Rev Vaccines.* 2018 Jun;17(6):525-545.
37. Leach CN, Johnson HN. Human rabies, with special reference to virus distribution and titer. *Amer J Trop Med.* 1940 March; s1-20(2):335-340.
38. Koprowski H, Cox HR. Studies on chick embryo adapted rabies virus; culture characteristics and pathogenicity. *J Immunol.* 1948;60(4):533–554.
39. Remlinger P, Bailly J, Hadji A. Contribution à l'étude du virus rabique Flury. *Bull Acad Natl Med.* 1953 Jun 9-23;137(21-23):340-2.
40. Koprowski H. Biological modification of rabies virus as a result of its adaptation to chicks and developing chick embryos. *Bull World Health Organ.* 1954;10(5):709-24.
41. Koprowski H, Black J. Studies on chick-embryo-adapted rabies virus. VII. Immunological responses of animals to vaccination with high egg passage Flury strain. *J Immunol.* 1954 Jun;72(6):503-10.
42. Schwab MP, Fox JP, Conwell DP, Robinson TA. Avianized rabies virus vaccination in man. *Bull World Health Organ.* 1954;10(5):823-35.
43. Atanasiu P, Bahmanyar M, Baltazard M, et al. Rabies neutralizing antibody response to different schedules of serum and vaccine inoculations in non-exposed persons. *Bull World Health Organ.* 1956;14(4):593-611.
44. Dean DJ, Evans WM, Thompson WR. Studies on the low egg passage Flury strain of modified live rabies virus produced in embryonating chicken eggs and tissue culture. *Am J Vet Res.* 1964 May;25:756-63.
45. Nicholson KG, Farrow PR, Bijok U, Barth R. Pre-exposure studies with purified chick embryo cell culture rabies vaccine and human diploid cell vaccine: serological and clinical responses in man. *Vaccine.* 1987 Sep;5(3):208-10.
46. Madhusudana SN, Tripathi KK. Post exposure studies with human diploid cell rabies vaccine and purified chick embryo cell vaccine: comparative serological responses in man. *Zentralbl Bakteriol.* 1989 Sep;271(3):345-50.
47. Wasi C, Chaiprasithikul P, Auewarakul P, Puthavathana P, Thongcharoen P, Trishnananda M. The abbreviated 2-1-1 schedule of purified chick embryo cell rabies vaccination for rabies postexposure treatment. *Southeast Asian J Trop Med Public Health.* 1993 Sep;24(3):461-6.
48. Suntharasamai P, Chaiprasithikul P, Wasi C, et al. A simplified and economical intradermal regimen of purified chick embryo cell rabies vaccine for postexposure prophylaxis. *Vaccine.* 1994 May;12(6):508-12.

49. Kulkarni R, Thatte U, Shinde V, Dharadhar S, Popova O, Vakil H. A comparison of the tolerability of two dilution volumes (0.5 mL and 1.0 mL) of a purified chick embryo cell rabies vaccine administered intramuscularly to healthy adult volunteers: A randomized, intraindividual, assessor-blind study. *Curr Ther Res Clin Exp.* 2004 Jan;65(1):47-56.
50. Briggs DJ, Banzhoff A, Nicolay U, et al. Antibody response of patients after postexposure rabies vaccination with small intradermal doses of purified chick embryo cell vaccine or purified Vero cell rabies vaccine. *Bull World Health Organ.* 2000;78(5):693-8.
51. Madhusudana SN, Anand NP, Shamsundar R. Economical multi-site intradermal regimen with purified chick embryo cell vaccine (Rabipur) prevents rabies in people bitten by confirmed rabid animals. *Int J Infect Dis.* 2002 Sep;6(3):210-4.
52. Kerdpanich P, Chanthavanich P, De Los Reyes MR, et al. Shortening intradermal rabies post-exposure prophylaxis regimens to 1 week: Results from a phase III clinical trial in children, adolescents and adults. *PLoS Negl Trop Dis.* 2018 Jun 6;12(6):e0006340.
53. Ma J, Wang H, Li J, et al. A randomized open-labeled study to demonstrate the non-inferiority of purified chick-embryo cell rabies vaccine administered in the Zagreb regimen (2-1-1) compared with the Essen regimen in Chinese adults. *Hum Vaccin Immunother.* 2014;10(10):2805-12.
54. Mahendra BJ, Narayana DA, Agarkhedkar S, et al. Comparative study on the immunogenicity and safety of a purified chick embryo cell rabies vaccine (PCECV) administered according to two different simulated post exposure intramuscular regimens (Zagreb versus Essen). *Hum Vaccin Immunother.* 2015;11(2):428-34.
55. Tanterdtham S, Chairasithikul P, Yuthavong K, Wasi C. Follow-up of protective antibody level after post-exposure vaccination with purified tissue culture rabies vaccine (PCEC) small doses intradermally. *J Med Assoc Thai.* 1991 Nov;74(11):498-501.
56. Ravish HS, Vijayashankar V, Madhusudana SN, et al. Safety and Immunogenicity of purified chick embryo cell rabies vaccine (VaxiRab N) administered intradermally as post exposure prophylaxis. *Hum Vaccin Immunother.* 2014;10(8):2433-7.
57. Madhusudana SN, Anand NP. Response to purified chick embryo cell rabies vaccine administered intradermally for post-exposure prophylaxis. *Natl Med J India.* 1997 May-Jun;10(3):115-6.
58. Madhusudana SN, Sanjay TV, Mahendra BJ, Suja MS. Simulated post-exposure rabies vaccination with purified chick embryo cell vaccine using a modified Thai Red Cross regimen. *Int J Infect Dis.* 2004 May;8(3):175-9.
59. Madhusudana SN, Anand NP, Shamsundar R. Evaluation of two intradermal vaccination regimens using purified chick embryo cell vaccine for post-exposure prophylaxis of rabies. *Natl Med J India.* 2001 May-Jun;14(3):145-7.
60. Quiambao BP, Dimaano EM, Ambas C, et al. Reducing the cost of post-exposure rabies prophylaxis: efficacy of 0.1 ml PCEC rabies vaccine administered intradermally using the Thai Red Cross post-exposure regimen in patients severely exposed to laboratory-confirmed rabid animals. *Vaccine.* 2005 Feb 25;23(14):1709-14.
61. Chhabra M, Ichhpujani RL, Bhardwaj M, Tiwari KN, Panda RC, Lal S. Safety and immunogenicity of the intradermal Thai red cross (2-2-2-0-1-1) post exposure vaccination regimen in the Indian population using purified chick embryo cell rabies vaccine. *Indian J Med Microbiol.* 2005 Jan;23(1):24-8.
62. Sudarshan MK, Madhusudana SN, Mahendra BJ, et al. Evaluation of a new five-injection, two-site, intradermal schedule for purified chick embryo cell rabies vaccine: A randomized, open-label, active-controlled trial in healthy adult volunteers in India. *Curr Ther Res Clin Exp.* 2005 Jul;66(4):323-34.
63. Madhusudana SN, Sanjay TV, Mahendra BJ, et al. Comparison of safety and immunogenicity of purified chick embryo cell rabies vaccine (PCECV) and purified vero cell rabies vaccine (PVRV) using the Thai Red Cross intradermal regimen at a dose of 0.1 ML. *Hum Vaccin.* 2006 Sep-Oct;2(5):200-4.
64. Sudarshan MK, Gangaboraiah B, Ravish HS, Narayana DH. Assessing the relationship between antigenicity and immunogenicity of human rabies vaccines when administered by intradermal route: results of a metaanalysis. *Hum Vaccin.* 2010 Jul;6(7):562-5.
65. Ashwathnarayana DH, Madhusudana SN, Sampath G, et al. A comparative study on the safety and immunogenicity of Purified duck embryo vaccine [corrected] (PDEV, Vaxirab) with purified chick embryo cell vaccine (PCEC, Rabipur) and purified vero cell rabies vaccine (PVRV, Verorab). *Vaccine.* 2009 Dec 10;28(1):148-51.

66. Sudarshan MK, Madhusudana SN, Mahendra BJ, et al. Boosting effect of purified chick embryo cell rabies vaccine using the intradermal route in persons previously immunized by the intramuscular route or vice versa. *Natl Med J India*. 2006 Jul-Aug;19(4):192-4.
67. Ravish HS, Sudarshan MK, Madhusudana SN, et al. Assessing safety and immunogenicity of post-exposure prophylaxis following interchangeability of rabies vaccines in humans. *Hum Vaccin Immunother*. 2014;10(5):1354-8.
68. Sudarshan MK, Narayana DH, Madhusudana SN, et al. Evaluation of a one week intradermal regimen for rabies post-exposure prophylaxis: results of a randomized, open label, active-controlled trial in healthy adult volunteers in India. *Hum Vaccin Immunother*. 2012 Aug;8(8):1077-81.
69. Briggs DJ, Dreesen DW, Nicolay U, et al. Purified Chick Embryo Cell Culture Rabies Vaccine: interchangeability with Human Diploid Cell Culture Rabies Vaccine and comparison of one versus two-dose post-exposure booster regimen for previously immunized persons. *Vaccine*. 2000 Dec 8;19(9-10):1055-60.
70. Malerczyk C, Briggs DJ, Dreesen DW, Banzhoff A. Duration of immunity: an anamnestic response 14 years after rabies vaccination with purified chick embryo cell rabies vaccine. *J Travel Med* 2007;14:63-4.
71. Sudarshan MK, Giri MS, Mahendra BJ, et al. Assessing the safety of post-exposure rabies immunization in pregnancy. *Hum Vaccin* 2007;3:87-9.
72. Li R, Li Y, Wen S, et al. Immunogenicity and safety of purified chick-embryo cell rabies vaccine under Zagreb 2-1-1 or 5-dose Essen regimen in Chinese children 6 to 17 years old and adults over 50 years: a randomized open-label study. *Hum Vaccin Immunother*. 2015;11(2):435-42.
73. Lumbiganon P, Chaiprasithikul P, Sookpranee T, Paholpak S, Wasi C. Pre-exposure vaccination with purified chick embryo cell rabies vaccines in children. *Asian Pac J Allergy Immunol*. 1989 Dec;7(2):99-101.
74. Shanbag P, Shah N, Kulkarni M, et al. Protecting Indian schoolchildren against rabies: pre-exposure vaccination with purified chick embryo cell vaccine (PCECV) or purified verocell rabies vaccine (PVRV). *Hum Vaccin*. 2008 Sep-Oct;4(5):365-9.
75. Pengsaa K, Limkittikul K, Sabchareon A, et al. A three-year clinical study on immunogenicity, safety, and booster response of purified chick embryo cell rabies vaccine administered intramuscularly or intradermally to 12- to 18-month-old Thai children, concomitantly with Japanese encephalitis vaccine. *Pediatr Infect Dis J*. 2009 Apr;28(4):335-7.
76. Alberer M, Burchard G, Jelinek T, et al. Co-administration of a meningococcal glycoconjugate ACWY vaccine with travel vaccines: a randomized, open-label, multi-center study. *Travel Med Infect Dis*. 2014 Sep-Oct;12(5):485-93.
77. Sampath G, Parikh S, Sangram P, Briggs DJ. Rabies post-exposure prophylaxis in malnourished children exposed to suspect rabid animals. *Vaccine* 2005;23:1102-5.
78. Sirikwin S, Likanonsakul S, Waradejwinyoo S, et al. Antibody response to an eight-site intradermal rabies vaccination in patients infected with Human Immunodeficiency Virus. *Vaccine*. 2009 Jul 9;27(32):4350-4.
79. Sampath G, Banzhoff A, Deshpande A, et al. Comparison of the immunogenicity and safety of the purified chick embryo cell rabies vaccine manufactured in India and Germany: A randomized, single blind, multicentre, phase IV clinical study. *Hum Vaccin Immunother*. 2017 Jul 3;13(7):1531-1538.
80. Malerczyk C, Selhorst T, Tordo N, et al. Antibodies induced by vaccination with purified chick embryo cell culture vaccine (PCECV) cross-neutralize non-classical bat lyssavirus strains. *Vaccine* 2009;27:5320-5.
81. Mani RS, Dovih DP, Ashwini MA, et al. Serological Evidence of Lyssavirus Infection among Bats in Nagaland, a North-Eastern State in India. *Epidemiol Infect*. 2017 Jun;145(8):1635-1641.
82. Abela-Ridder B, Balogh de K, Kessels JA, Dieuzy-Labaye I, Torres G. Global rabies control: the role of international organisations and the Global Strategic Plan to eliminate dog-mediated human rabies. *Rev Sci Tech*. 2018 Aug;37(2):741-749.
83. Tantawichien T, Rupprecht CE. Modern biologics for rabies prophylaxis and the elimination of human cases mediated by dogs. *Expert Opin Biol Ther*. 2020 Nov;20(11):1347-1359.
84. Rupprecht CE, Abela-Ridder B, Abila R, et al. Towards rabies elimination in the Asia-Pacific region: From theory to practice. *Biologicals*. 2020 Mar;64:83-95.

85. Rupprecht CE, Salahuddin N. Current status of human rabies prevention: remaining barriers to global biologics accessibility and disease elimination. *Expert Rev Vaccines*. 2019 Jun;18(6):629-640.
86. SAGE Working Group on Rabies Vaccines and Immunoglobulins and the World Health Organization Secretariat, 2017. Background Paper: Proposed Revision of the Policy on Rabies Vaccines and Rabies Immunoglobulins. Available at: http://www.who.int/immunization/sage/meetings/2017/october/1_Background_paper_WG_RABIES_final.pdf.
87. Cantaert T, Borand L, Kergoat L, et al. A 1-week intradermal dose-sparing regimen for rabies post-exposure prophylaxis (RESIST-2): an observational cohort study. *Lancet Infect Dis*. 2019 Dec;19(12):1355-1362.
88. Kessels JA, Recuenco S, Navarro-Vela AM, et al. Pre-exposure rabies prophylaxis: a systematic review. *Bull World Health Organ*. 2017 Mar 1;95(3):210-219C.
89. Sreenivasan N, Li A, Shiferaw M, et al. Overview of rabies post-exposure prophylaxis access, procurement and distribution in selected countries in Asia and Africa, 2017-2018. *Vaccine*. 2019 Oct 3;37 Suppl 1:A6-A13.
90. Taylor E, Banyard AC, Bourhy H, et al. Avoiding preventable deaths: The scourge of counterfeit rabies vaccines. *Vaccine*. 2019 Apr 17;37(17):2285-2287.

National Guidelines for Rabies Prophylaxis

Summary of Vaccination Schedule

The summary of vaccination schedule as per route is as under :

Type of Prophylaxis	Route of Administration	Dose of Vaccine	Day of Dose	No. of injections	Total No. of Per Visit	Site of Injection Visits
Post Exposure Prophylaxis	Intra Dermal	0.1 ml per dose	Day 0, 3, 7 and 28	2	4	Adult: Deltoid Muscle Infants and Small Children: Anterolateral Thigh
	Intra Muscular	1 entire vaccine vial	Day 0, 3, 7, 14 and 28	1	5	
Pre Exposure Prophylaxis	Intra Dermal	0.1 ml per dose	Day 0, 7, and booster on either day 21 or 28	1	3	
	Intra Muscular	1 entire vaccine vial	Day 0, 7, and booster on either day 21 or 28	1	3	
Re-exposure	Intra Dermal	0.1 ml per dose	Day 0 & 3	1	2	
	Intra Muscular	1 entire vaccine vial	Day 0 & 3	1	2	